

Earthworm species in *Musa* spp. plantations in Brazil and worldwide

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Abstract

Bananas and plantains are major commodity/food crops that represent an important habitat for earthworms, although so far, no review is available on earthworm communities associated with banana/plantain crops worldwide. The Vale do Ribeira region is among the largest banana producing areas in Brazil, but little is known of the earthworms living there. Hence, the present study assessed earthworm populations and species in three banana plantations and adjacent Atlantic forest fragments along the Ribeira de Iguape River using standard (hand sorting) methodologies. Furthermore, we review earthworm populations reported in banana/plantain plantations worldwide. Only two species (*Pontoscolex corethrurus*, *Amyntas gracilis*) belonging to two families (Rhinodrilidae, Megascolecidae) were found in the Ribeira River valley, occurring concurrently. Abundance was low (< 13 indiv. m^{-2}) compared with other banana plantations worldwide, that frequently surpassed 100 indiv. m^{-2} . More than 70 studies reported earthworms from >200 banana plantations in 28 countries, and mean species richness was 2.7 per site, ranging from 1 to 10 species. Exotics predominated in most sites and *P. corethrurus* was the most prevalent species encountered. Overall, more than 104 species from 10 families were reported, with around 61 native and 43 exotic widespread species, mainly of the Megascolecidae, Lumbricidae and Acanthodrilidae families. Richness was highest in India (27 spp.) and the Canary Islands (25 spp.), but native species dominated only in a few countries and sites, while exotics were prevalent especially in island countries and Brazil. Lower-input practices appear to be important for earthworm communities and banana plantations can have large earthworm populations in

some cases, which may be contributing to soil processes and plant production, topics that deserve further attention. However, many important banana-producing countries have not yet been evaluated, so further work is warranted, both in terms of applied ecology and biodiversity.

Keywords

Annelida, banana, biodiversity, Oligochaeta, plantain, *Pontoscolex corethrurus*

Introduction

Bananas and plantains are large, perennial herbs belonging to the genus *Musa*, that evolved in Indochina and Southeast Asia, but with major secondary diversification in Africa, India and the Caribbean (Price 1995). Bananas are a major commodity, occupying over 6 million ha (FAO 2018) and representing an important contribution to the economy of many developing countries worldwide (OECD/FAO 2019). Plantains resemble bananas, but are generally longer, have more starch and are mostly eaten cooked, rather than raw (like the bananas). They are a major staple crop in several African, Asian, Pacific, Latin American and Caribbean countries (Price 1995; Norgrove and Hauser 2014). In 2018, the six main banana producers (total production) were India, China, Indonesia, Brazil, Ecuador and the Philippines, while the six countries with the greatest surface area devoted to banana production were India (884,000 ha), Tanzania (490,701 ha), Philippines (484,247 ha), Rwanda (464,321 ha), Brazil (449,284 ha) and China (383,216 ha) (FAO 2018). India accounts for around 24% of global production and Brazil around 5% (FAO 2018), while the whole of Latin America and the Caribbean (LAC) region account for around 25% of the world's banana production (OECD/FAO 2019).

Throughout much of LAC, bananas and plantains are still cultivated at the subsistence level, often in agroforestry systems (Harvey and Villalobos 2007; Malézieux et al. 2009; Paul et al. 2015; Coelho 2017; Garcia et al. 2017; Salazar-Díaz and Tixier 2017). However, commercial plantations are also widespread, occupying large monoculture areas, particularly in warmer, wetter regions of the tropics (Campbell 2018; Yahia 2019). In Brazil, most of the area devoted to banana cultivation lies within the Atlantic Rainforest biome, a highly threatened hotspot of biodiversity (Myers et al. 2000). In fact, much of the banana and plantain cultivation worldwide is performed in wetter tropical climates, and frequently close to rainforest ecosystems, where they may represent a potential hazard to biodiversity conservation. In commercial plantations, conventional production practices are adopted, including frequent herbicide use to control weeds, fumigation to control fungal diseases (particularly *Fusarium* and *Pythium*) and root nematode infestation, as well as Sigatoka (Marin et al. 2003; Cordeiro et al. 2004; Gasparotto et al. 2006), although some resistant varieties for the latter are already available (Timm et al. 2016; Dale et al. 2017). These practices may have important negative impacts on earthworm populations (da Silva et al. 2006; Baretta et al. 2011), despite the high amounts of litter inputs, which represent C (food) sources for

soil biota, and protection from soil erosion (Lombardi Neto and Moldenhauer 1992). Worldwide, however, little is known of the soil biota inhabiting banana plantations, and so far, there has not been an overview of true soil-inhabiting animals in banana plantations worldwide.

Earthworms are essential service providers for terrestrial ecosystems (Lavelle et al. 2006). Their activity, generating galleries and casts, contributes to formation and maintenance of soil structure (Lavelle 1997; Capowiez et al. 2012), increasing porosity, infiltration and water retention (Fiuza et al. 2012), as well as re-distribution and breakdown of soil organic matter (Brown et al. 2000). However, earthworms are sensitive to land use and management, and can be used as soil quality and management as well as environmental bioindicators (Brown and Domínguez 2010; Bartz et al. 2013; Bünemann et al. 2018). Brazil is home to more than 300 described earthworm species (Brown et al. 2013), but practically nothing is known of the species and populations inhabiting banana plantations in the country.

The Vale do Ribeira region, located in northeastern Paraná State and southern São Paulo State, has extensive areas (over 36,000 hectares; ABAVAR 2015) devoted to banana cultivation (Bueno 2003). In this region, banana fields are normally surrounded by Atlantic forest fragments (Cordeiro et al. 2017), that have been reduced to around 12% of their original surface area (Ribeiro et al. 2009). Although frequently disturbed with various management practices, banana plantations are perennial crops that could provide adequate habitats for the establishment of native earthworm species, especially when Atlantic forest fragments occur surrounding banana cropping areas (Cordeiro et al. 2017). However, little is known about the effects of banana crops on abundance and diversity of earthworm species, and the occurrence of these invertebrates in Atlantic forest fragments in the Ribeira valley region. Furthermore, little is known of the presence of native and exotic earthworm species in banana and plantain fields worldwide. Hence, the present study was undertaken to assess earthworm populations in banana plantations and native forest fragments in the Ribeira de Iguape River valley in the State of São Paulo, and evaluate earthworm communities (abundance, biomass, species composition) associated with banana and plantain crops worldwide.

Material and methods

Study sites in the Ribeira de Iguape River valley

Three counties in the lower Ribeira River valley, all of them in the State of São Paulo were selected for this study: Eldorado, Sete Barras and Registro (Fig. 1). The climate in Sete Barras and Registro is rainy tropical (Af-type according to Köppen), with mean rainfall greater than 60 mm in the driest month. In Eldorado, climate is Köppen Am tropical, with rainfall less than 60 mm in the driest month. The average annual rainfall for all counties ranges from 1500 to 1600 mm (CEPAGRI 2018; CIIAGRO 2018), with the highest concentration of rains occurring from January to March. The mean

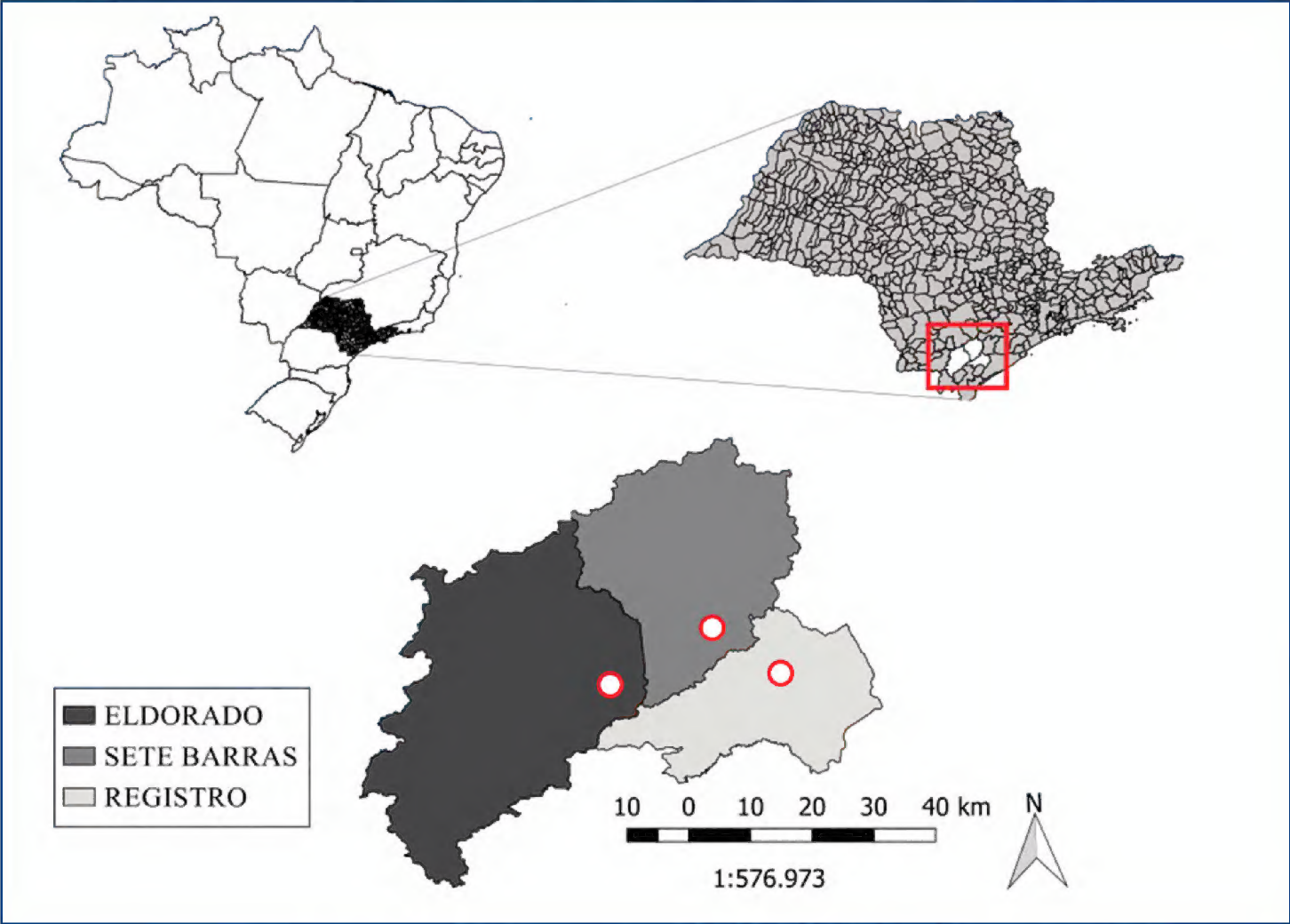


Figure 1. Location of the counties sampled in the Ribeira de Iguape River valley, São Paulo State, Brazil.

Table 1. Land use system, watershed number (WN), age of the land use, geographic coordinates and soil types according to FAO classification (IUSS/WRB 2015) of the sites evaluated in each county of the Ribeira de Iguape River Valley, São Paulo, Brazil.

Site	County	System	WN ¹	Age (yrs)	Latitude, Longitude	Soil types
1	Eldorado	Banana	344	50	24°29'35"S, 48°02'10"W	Cambisols
2	Eldorado	Atlantic forest	344	> 50	24°30'09"S, 48°02'30"W	Cambisols
3	Sete Barras	Banana	422	15	24°23'34"S, 47°53'51"W	Cambisols
4	Sete Barras	Atlantic forest	422	> 50	24°23'30"S, 47°53'22"W	Cambisols
5	Registro	Banana	379	40	24°26'56"S, 47°49'41"W	Cambisols / Histosols
6	Registro	Atlantic forest	389	45	24°26'47"S, 47°49'23"W	Cambisols / Histosols

¹Official cartographic number for the watershed.

annual temperature ranges from 23.9 to 24.3 °C, with the lowest temperature (13 °C) in July and highest (34.2 °C) in February. Soils in the valley originate from sedimentary, metabasic and amphibolic rocks (Oliveira et al. 2002), with high natural fertility (calcium, magnesium, potassium, and phosphorus content) and high organic matter levels, due to seasonal river floods that deposit alluvial material. Soil texture varies from loam to clay. The areas chosen in the three counties are characterized by smaller watersheds that flow into the Ribeira River with banana crops on the high ground level and Atlantic forest sites (control sites) in advanced stages of regeneration close to the Ribeira River. General characteristics of the areas are given in Table 1.

Earthworm sampling

Earthworms were collected using an adaptation of the standard sampling method proposed by the Tropical Soil Biology and Fertility (TSBF) Programme (Anderson and Ingram 1993). In each area 10 samples (25 × 25 cm square to 20 cm depth) were taken, divided into 2 equally-numbered transects with samples every 20 m. Distance between transects was ca 10 m. Earthworms were hand-sorted from the soil in the field and fixed in 80% alcohol. In the laboratory, earthworms were identified to species or family level (juveniles) using taxonomic keys (Michaelsen 1900; Righi 1990; Blakemore 2002). The material was deposited in the Fritz Müller Oligochaete collection (COFM) at Embrapa Forestry in Colombo, Brazil. The earthworm data obtained were used to determine the total species abundances (no. individuals and fresh mass m⁻²) and richness, per site and land use (banana, forest).

Literature review

Both the common and scientific names of banana were used for a bibliographic search online using the keywords for bananas and plantains in English, Portuguese, French and Spanish: *Musa* (genus), *Musa acuminata*, *Musa balbisiana*, banana, banane, banano, plátano and plantain. These were then crossed with the common names of earthworms in these languages: earthworms, minhoca, oligochaeta, oligoqueta, vers de terre and lombriz de tierra. Online scientific databases Web of Science, Science Direct, Scielo, google academic and the Base de Dados de Teses e Dissertações (BDTD – Thesis and Dissertation Database) of Brazil were consulted. All the resulting publications were consulted and those containing data on earthworm abundance (density and/or biomass) or species identification were selected and these data extracted, as well as information on sampling sites (counties, countries, management practices of the plantations). Earthworm species were separated into different families and into native or exotic to the region of occurrence, and species richness per site and for each group (native, exotic), when available. Although we treated bananas and plantains separately when possible, for most of the analysis we considered them together, since not all publications provided details regarding the types of bananas cultivated, and even plantains are often called ‘bananas.’ Details on the species and management data obtained and presented in this paper are available for download online from the open access repository Mendeley Data at <http://dx.doi.org/10.17632/p8ywsnj8c5.1> (Cremonesi et al. 2020).

Data treatment

Quantitative data on the earthworm abundance and biomass obtained from the literature and from the present study were treated as follows. Means of earthworm abundance (no. individuals m⁻²) and biomass (fresh mass in gm⁻²) were calculated per sampling site (plantation), using data from the present study. When quantitative data from the litera-

ture was available for the individual site, it was used as is. When only means for several plantations in the same general location were provided, these were also used. As the interest of the present study was more at the spatial (site-level) rather than the temporal scale, when samples were taken on multiple occasions, and individual means per sampling date were not available, overall means were used. When taken in wet and dry seasons, both values were used as an interval of abundance and biomass (when measured).

Results and discussion

Specimens examined from the Ribeira de Iguape River valley sites

Family Rhinodrilidae

Pontoscolex (Pontoscolex) corethrurus (Müller, 1857)

COFMBRSP0231, 1 individual in Atlantic Forest, HMN 389, Registro – SP (24°26'16.85"S, 47°49'31.71"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0232, 2 individuals in Atlantic Forest, HMN 389, Registro – SP (24°26'16.82"S, 47°49'31.71"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0233, 2 individuals in Atlantic Forest, HMN 389, Registro – SP (24°26'16.28"S, 47°49'32.52"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0235, 2 individuals in Atlantic Forest, HMN 389, Registro – SP (24°26'15.71"S, 47°49'33.32"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0236, 1 individual in Atlantic Forest, HMN 389, Registro – SP (24°26'14.57"S, 47°49'35.35"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0238, 2 individuals in banana field, HMN 379, Registro – SP (24°26'54.25"S, 47°49'38.12"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0239, 1 individual in banana field, HMN 379, Registro – SP (24°26'54.81"S, 47°49'39.41"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0240, 1 individual in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'44.43"S, 47°55'11.56"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0241, 1 individual in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'44.46"S, 47°55'11.49"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0242, 2 individuals in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'43.79"S, 47°55'24.53"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0244, 1 individual in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'43.93"S, 47°55'10.17"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0245, 3 individuals in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'44.33"S, 47°55'09.65"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0248, 1 individual in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'44.90"S, 47°55'08.92"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0249, 1 individual in banana field, HMN 422, Sete Barras – SP (24°23'38.61"S, 47°55'23.49"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0251, 1 individual in banana field, HMN 422, Sete Barras – SP (24°23'43.01"S, 47°55'24.52"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0252, 3 individuals in banana field, HMN 422, Sete Barras – SP

(24°23'42.54"S, 47°55'25.32"W), 2019, M. Cremonesi, A. Santos colls. COFMBR-SP0253, 1 individual in Atlantic Forest, HMN 344, Eldorado – SP (24°29'57.34"S, 48°02'41.68"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0255, 1 individual in Atlantic Forest, HMN 344, Eldorado – SP (24°29'55.69"S, 48°02'42.15"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0256, 2 individuals in banana field, HMN 344, Eldorado – SP (24°29'36.89"S, 48°02'09.43"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0258, 2 individuals in banana field, HMN 344, Eldorado – SP (24°29'37.11"S, 48°02'10.84"W), 2019, M. Cremonesi, A. Santos colls.

Rhinodrilidae juveniles. COFMBRSP0246, 1 individual in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'44.33"S, 47°55'09.65"W), 2019, M. Cremonesi, A. Santos colls.

Family Megascolecidae

Amyntas gracilis (Kinberg, 1867)

COFMBRSP0237, 1 individual in banana field, HMN 379, Registro – SP (24°26'54.25"S, 47°49'38.22"W), 2019, M. Cremonesi, A. Santos colls. COFMBR-SP0250, 3 individuals in banana field, HMN 422, Sete Barras – SP (24°23'38.61"S, 47°55'23.49"W), 2019, M. Cremonesi, A. Santos colls.

Megascolecidae juveniles. COFMBRSP0234, 1 individual in Atlantic Forest, HMN 389, Registro – SP (24°26'16.28"S, 47°49'32.52"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0243, 1 individual in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'44.06"S, 47°55'10.35"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0247, 1 individual in Atlantic Forest, HMN 422, Sete Barras – SP (24°23'44.33"S, 47°55'09.65"W), 2019, M. Cremonesi, A. Santos colls. COFMBR-SP0254, 1 individual in Atlantic Forest, HMN 344, Eldorado – SP (24°29'56.60"S, 48°02'42.23"W), 2019, M. Cremonesi, A. Santos colls. COFMBRSP0257, 1 individual in banana field, HMN 344, Eldorado – SP (24°29'36.89"S, 48°02'09.43"W), 2019, M. Cremonesi, A. Santos colls.

Earthworm populations in the Ribeira River valley and other sites in Brazil

Only two earthworm species belonging to two families (Rhinodrilidae, Megascolecidae) were found at the six sampling sites in the three counties (Table 2): *Pontoscolex* (*Pontoscolex*) *corethrurus* and *Amyntas gracilis*, both considered peregrine/exotic in southern Brazil (Brown et al. 2006). *Pontoscolex corethrurus* may have originated in the Guyana shield area (Righi 1984), and *A. gracilis* may be native to China (Blakemore 2002). The former species was found living in all sites, while the latter was found in both banana plantations and native forest in Sete Barras and in banana plantations in Registro. At the other sites, only juveniles of the Megascolecidae family were found. These were most likely *A. gracilis* as well, but could not be identified to species level. Maximum richness found per site was similar in banana crops and Atlantic forest fragments (two spp. in each land use), but with some variation between sites (Table 2).

Table 2. Earthworm families, species, and richness in banana plantations and Atlantic Forest remnants, in three counties of the Ribeira de Iguape River valley (Eldorado, Sete Barras, Registro). + means presence and – means absence.

Earthworm family and species	Eldorado		Sete Barras		Registro	
	Banana	Atlantic Forest	Banana	Atlantic Forest	Banana	Atlantic Forest
Megascolecidae						
<i>Amyntas gracilis</i>	–	–	+	+	+	–
Megascolecidae juveniles	+	+	–	+	–	+
Rhinodrilidae						
<i>Pontoscolex corethrurus</i>	+	+	+	+	+	+
Rhinodrilidae juveniles	–	–	–	+	–	–
Species Richness	2	2	2	≥2	2	2

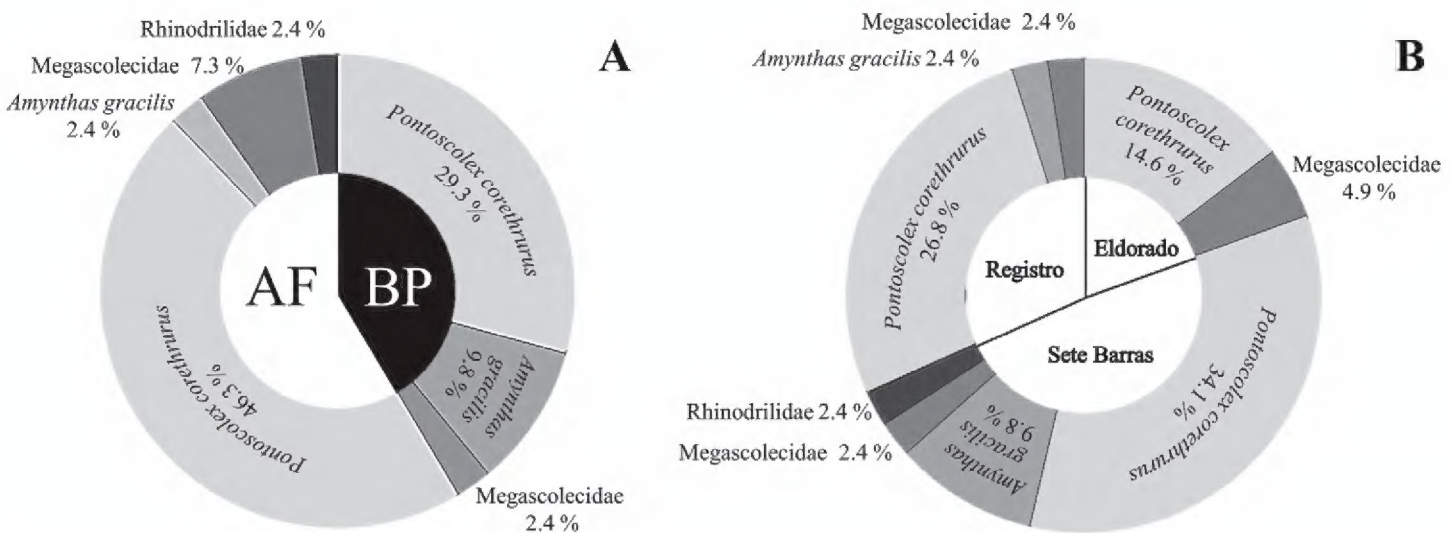


Figure 2. Frequency of earthworm species (% of total individuals collected) in each ecosystem sampled (A) in Atlantic Forest (AF) and banana plantations (BP) and by counties (B).

Most of the individuals collected (76% of the total) were of *P. corethrurus*, representing 29% of the total abundance in banana crop sites and 46% in Atlantic forest fragments (Fig. 2). *Amyntas gracilis*, although not occurring in all areas, accounted for 12% of all individuals sampled, of which 10% were found in banana crops but only 2% in Atlantic forests. Rhinodrilidae juveniles represented only 2% of the earthworms found, and occurred only in the Atlantic forest, while Megascolecidae juveniles represented 10% of all earthworms, and were often found in Atlantic forest fragments. Both species are widespread in Brazil (Brown et al. 2006), especially in agricultural and disturbed ecosystems, and display relatively high tolerance to a range of abiotic/biotic conditions, which have allowed these species to spread throughout most of the tropics and subtropics worldwide (Brown et al. 2006; González et al. 2006; Taheri et al. 2018). They have also been recommended as indicators of soil quality in agroecosystems and of disturbance in natural landscapes (Nunes et al. 2007; Fernandes et al. 2010).

The predominance of *P. corethrurus* in both native forest and banana plantations of the Ribeira River valley indicate that non-native species have extensively colonized disturbed soils of this region. Nonetheless, this potentially widespread occurrence of exotics should be further evaluated both regionally and nationally, in order to better determine the extent of this phenomenon as well as its possible causes.

Mean overall abundance and biomass of earthworms found in the three sites studied here (6 to 13 indiv. m⁻² and 2.5 to 9 g m⁻²) tended to be quite low compared with others observed overall in Brazil (21 to 459 indiv. m⁻² and 3.1 to 177.4 g m⁻²; see Table 3). At sites near the Ribeira River valley in the neighboring state of Paraná (Römbke et al. 2009; Maschio et al. 2010), and within the Ribeira River watershed in the nearby Turvo River valley (a tributary of the Ribeira River; Brown et al. 2009), both abundance and biomass were generally much higher (Table 3), even though the predominant earthworm species was the same (*P. corethrurus*). This is probably due to the less intensive and more traditional agroforestry management practices used in these sites, including slashing and mulching, as well as the presence of other trees, particularly atmospheric N₂-fixing leguminous trees, and the absence of or lower pesticide use (Brown et al. 2009; Römbke et al. 2009). These practices may benefit earthworm populations, particularly *P. corethrurus*, as observed comparing a mulched and non-mulched plantation in Antonina, where earthworm abundance was ~13 times higher with mulching (Maschio et al. 2010). Reasons for the lower values found in the Ribeira River valley sites may be due to the more intensive management practices typical of commercial banana plantations in the region, including insecticide and nematicide applications, which may reduce earthworm populations (Clermont-Dauphin et al. 2004).

Earthworm communities in banana plantations worldwide

More than 70 studies were found from 28 countries with data on earthworms in banana and plantain fields (Tables 3, 4, 5). Of these studies, 49 had species data (Table 4; see also full dataset in Cremonesi et al. 2020), coming from ≥ 210 sites (Table 5), of which most were in the Spanish Canary Islands (*N* = 77), mainly due to the intensive sampling efforts of Talavera in Tenerife (Talavera 1992a). Interestingly, two of the major banana-producing countries in terms of area were not represented (Tanzania, Rwanda), and in China (another important producer), only one study reported earthworms from a single site (Sun et al. 2012). Plantain banana fields were sampled in only 22 locations (10% of total) in four countries (Colombia, Cameroon, Ivory Coast and Ecuador; Tondoh 2007; Norgrove et al. 2011; Avilés 2017; Feijoo et al. 2018), and involved traditional management practices, rather than conventional cultivation. Most of the fields evaluated were banana plantations, and only in Ecuador were mixed banana/plantain fields evaluated (Avilés 2017).

Overall, ≥104 earthworm species from 10 earthworm families were recorded from banana/plantain fields worldwide, of which around 61 (59%) were native and 43 exotic to the sampling sites (Table 5). Estimating these numbers is difficult due to insufficient taxonomic resolution in some samples, as well as the uncertain origin of some widespread anthropochores (peregrines transported by humans), particularly in the Megascolecidae family (Blakemore 2002). Highest species richness (27) was observed overall in India, where most of the species found were native (74%). High proportions of native species were also observed in Ivory Coast, Madagascar, and Uganda (possibly 100%) as well as Cameroon (75%), but were lower in Mexico (58%) and Colombia

Table 3. Earthworm abundance and biomass found in banana plantations worldwide, and the predominant species encountered (when available).

Country	Location	Abundance (indiv. m ⁻²)	Biomass (g m ⁻²)	Predominant species	References
Brazil	Antonina (Monoculture)	71	35.1	<i>P. corethrurus</i>	Römbke et al. (2009)
		221	95.7	<i>P. corethrurus</i>	
		86	23.8	<i>P. corethrurus</i>	
	Antonina (Agroforestry)	173	77.1	<i>P. corethrurus</i>	Maschio et al. (2010)
		338	69.6	<i>P. corethrurus</i>	
		117	43.5	<i>P. corethrurus</i>	
		21 ^a	3.1 ^b	<i>P. corethrurus</i>	
		293 ^a	34.9 ^b	<i>P. corethrurus</i>	
	Adrianópolis (Agroforestry)	211–413 ^c	37–71.2 ^c	<i>P. corethrurus</i>	Brown et al. (2009)
	Barra do Turvo (Agroforestry)	99–176 ^c	11.2–17.3 ^c	<i>P. corethrurus</i>	
		229–459 ^c	48.3–117.4 ^c	<i>P. corethrurus</i>	
	Casimiro de Abreu	~205–440 ^c	–	NA	Quintero (2010)
	Paraty	167	–	NA	Correia et al. (2001)
	Eldorado	8	3.9	<i>P. corethrurus</i>	This study
	Sete Barras	13	9.0	<i>P. corethrurus</i>	
	Registro	6	2.5	<i>P. corethrurus</i>	
Cameroon	Mbalmayo Forest Reserve	70	–	<i>Legonodrilus</i> sp. nov. 1, <i>Eminoscolex lamani</i>	Norgrove et al. (2011)
		121	–	<i>Legonodrilus</i> sp. nov. 1, <i>Eminoscolex lamani</i>	
	Campo Ma'an	16–92 ^d	–	NA	Kanmegne (2004)
Colombia	Quindío (Armenia)	9–16 ^e	1.2–3.0 ^e	NA ^f	Molina and Feijoo (2016)
Costa Rica	Limón Province (Finca San Pablo)	83–812 ^g	–	NA	Agüero et al. (2002)
	Pueblo Nuevo de Villa Franca de Guácimo, Limón	29	6.2	NA	Cornwell (2014)
	Cahuita	350	144.6	<i>P. corethrurus</i>	Lapied and Lavelle (2003)
Guadeloupe (France)	Basse-Terre Andosols (mean of 23 sites)	88	23	NA	Clermont-Dauphin et al. (2004)
	Basse Terre Nitisols (mean of 11 sites)	54	17.5	NA	
	Capesterre-Belle-Eau (Gloria Bas)	168	27.6	<i>P. corethrurus</i>	Burac et al. (2018)
	Capesterre-Belle-Eau (Source)	288	42.2	<i>P. corethrurus</i>	
	Capesterre-Belle-Eau (Bergerie)	188	33.6	<i>P. corethrurus</i>	
	Baillif (Sextius)	336	112	<i>P. corethrurus</i>	
	Baillif (Grand Canon)	192	70.8	<i>P. corethrurus</i>	
	Saint-Claude (Saut d'Eau)	364	46	<i>P. corethrurus</i>	
Ecuador	Latacunga (La Maná)	168	–	NA	Avilés (2017)
		111	–	NA	
	Manabí (El Carmen)	78	–	NA	Figueroa (2019)
		37	–	NA	
	El Carmen (Cijádi)	0–145 ^h	–	NA	
	El Carmen (Nápoles)	34–144 ^h	–	NA	
	Santo Domingo de los Tsáchilas (Santa Patricia)	83–548 ^h	–	NA	
	Santo Domingo de los Tsáchilas (La Floresta)	22–150 ^h	–	NA	
India	West Tripura	16–656 ⁱ	4.8–453.6 ⁱ	<i>P. corethrurus</i>	Dhar and Chaudhuri (2018)
	Rajapalayam	116	48.8	<i>Lampito mauritii</i> , <i>Perionyx excavatus</i>	Mariappan et al. (2013)
Ivory Coast	Taabo (Lamto reservation)	186	8.5	<i>Reginaldia anomala</i>	Tondoh (1994, 2007)
Martinique (France)	Le Lorrain (Feugère)	244	67.6	<i>P. corethrurus</i>	Burac et al. (2018)
	Le Lorrain (Bellevue)	152	43.6	<i>P. corethrurus</i>	
	Le Lorrain (Limite)	52	26	<i>P. corethrurus</i>	
	L'Ajoupa-Bouillon (Allée Domergue 3)	148	49.6	<i>P. corethrurus</i>	
	Basse-Pointe (Fromager Rivière)	80	26	<i>P. corethrurus</i>	
	Basse-Pointe (Dantu Bas)	40	9	<i>P. corethrurus</i>	

Country	Location	Abundance (indiv. m ⁻²)	Biomass (g m ⁻²)	Predominant species	References
Mexico	Tabasco, Pablo L. Sidar	25	10	<i>P. corethrurus</i> , <i>Lavellodrilus</i> <i>bonampakensis</i>	Huerta et al. (2005)
	Tabasco, Teapa	116	20.8	<i>P. corethrurus</i> , <i>Drawida</i> <i>barwelli</i> , <i>Polypheretima</i> <i>elongata</i>	Geissen et al. (2009)
		117	11.8	<i>Balanteodrilus pearsei</i> , <i>Drawida barwelli</i>	
		94	40.4	<i>Balanteodrilus pearsei</i> , <i>Polypheretima elongata</i>	
		125	35.6	<i>P. corethrurus</i> , <i>Drawida</i> <i>barwelli</i>	
		25	8.8	<i>P. corethrurus</i> , <i>Lavellodrilus</i> <i>bonampakensis</i>	Huerta et al. (2007)
		~350	2.5	<i>Diploptrema murchiei</i>	Huerta et al. (2013)
		~350	9.3	<i>P. corethrurus</i>	
		~470	16.2	<i>P. corethrurus</i>	
		~100	11	<i>P. corethrurus</i>	
		~80	2.8	<i>P. corethrurus</i>	
		~125	0.8	<i>Dichogaster</i> sp.	
Nicaragua	León (Finca Cony)	150	–	NA	Hernández et al. (2015)
	León (Finca San Martín)	325	–	NA	
	León (Finca Santa Isabel)	50	–	NA	
	León (Finca El verdon)	65	–	NA	
	Possoltega (Finca San Joaquin)	150	–	NA	
	Possoltega (Finca Los Ángeles)	225	–	NA	
	Possoltega (Finca Maria de los Ángeles)	100	–	NA	
	Possoltega (Finca Montes Verdes)	125	–	NA	
Philippines	Davao (Sumitomo Fruits Corporation)	~85–175 ^j	–	NA	Fusilero et al. (2013)
		~75–215 ^j		<i>Metaphire cai</i>	
South Africa	Kwazulu-Natal (Eshowe)	1500 ^k	180	<i>Amyntas rodericensis</i> , <i>Amyntas minimus</i> , <i>P. corethrurus</i>	Dlamini and Haynes (2004)
Uganda	Kabanyolo University Farm	18–207 ^l	0.1–9.4 ^l	<i>Dichogaster</i> sp. 2, <i>Gordiodrilus</i> sp. 1	Block and Banage (1968)
	Mabira Forest reserve (1 yr old)	13	0.4	NA	Okwakol (1994)
	(2 yr old)	125	2.2	NA	
	(3 yr old)	131	1.3	NA	
	(5 yr old)	54	0.5	NA	
	(20 yr old)	154	4.2	NA	

^aEarthworm abundance values were corrected from Maschio et al. (2010) that reported earthworm numbers per sample and not per m².

^bBiomass values in g m⁻² are now included for this study. ^cMean of dry and wet season samplings, respectively. ^dMean abundance from eight sites, with four sampled in one year and the other four the subsequent year. ^eRange of abundance taken from eight replicate farms under four different management practices (totaling 32 plantations) in the Armenia region. ^fThe identification of the earthworm species collected overall in this study (not by plantation type) is published in Feijoo et al. (2018). ^gRange of abundance found under six weed control treatments (performed on same banana plantation) on five sampling dates; ^hRange of abundance found on six sampling dates in same plantation. ⁱRange of abundance and biomass found in three banana plantations. ^jRange of abundance and biomass found on sixteen sampling dates in same plantation. ^kMean of six banana plantations. ^lRange of abundance and biomass found on eight sampling dates in same plantation.

(53%). In these countries, many of the plantations were managed more traditionally, or using agroforestry, although the low number of sampling sites may also be responsible for these high values, particularly in the former countries. In fact, agroforestry systems had a total of 22 species from nine sites, while conventional production systems had only nine species from 13 sites. Nonetheless, because not enough information was provided in the publications on management practices (not reported in ≥150 sites;

Table 4. Earthworm species, richness and number of native and exotic species found in banana plantations under various management practices worldwide.

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
Bangladesh	Lalmonirhat District	NA	NA	<i>Lampito mauritii</i> , <i>Metaphire posthuma</i> , <i>Pontoscolex corethrurus</i>	3	2/1	Reynolds et al. (1995)
	Paget Parish	NA	NA	<i>Amyntas rodericensis</i>	1	0/1	Reynolds and Fragoso (2004)
Bermuda	Southampton Parish	NA	NA	<i>Amyntas lupetensis</i>	1	0/1	Reynolds and Fragoso (2004)
	Antonina, PR	Agroforestry	Monoculture	<i>Amyntas corticis</i> , <i>Pontoscolex corethrurus</i> , two other spp.	4	0/4	Römbke et al. (2009)
	Antonina, PR	Agroforestry	Monoculture	<i>Amyntas gracilis</i> , <i>Pontoscolex corethrurus</i> , two other spp.	4	0/4	Römbke et al. (2009)
	Antonina, PR	Agroforestry	Monoculture	<i>Dichogaster</i> spp., <i>Pontoscolex corethrurus</i>	4	0/4	Römbke et al. (2009)
	Antonina, PR	Agroforestry	Monoculture	<i>Ocnerodrilus occidentalis</i> , <i>Pontoscolex corethrurus</i> , two other spp.	4	0/4	Römbke et al. (2009)
	Antonina, PR	Agroforestry	Monoculture	<i>Pontoscolex corethrurus</i> , one other sp.	2	0/2	Römbke et al. (2009)
	Antonina, PR	NA	Polyculture	<i>Pontoscolex corethrurus</i> , one other sp.	2	0/2	Römbke et al. (2009)
	Antonina, PR	Agroforestry	Polyculture	<i>Dichogaster</i> sp., <i>Pontoscolex corethrurus</i> , and one unidentified sp.	3	?/2	Maschio et al. (2010)
	Antonina, PR	Agroforestry	Polyculture	<i>Pontoscolex corethrurus</i>	1	0/1	Maschio et al. (2010)
	Adrianópolis, PR	Agroforestry	Polyculture	<i>Amyntas gracilis</i> , <i>Pontoscolex corethrurus</i>	2	0/2	Brown et al. (2009)
	Barra do Turvo, SP	Agroforestry	Polyculture	<i>Amyntas gracilis</i> , <i>Pontoscolex corethrurus</i>	2	0/2	Brown et al. (2009)
	Barra do Turvo, SP	Agroforestry	Polyculture	<i>Amyntas gracilis</i> , <i>Dichogaster</i> sp., <i>Pontoscolex corethrurus</i>	3	0/3	Brown et al. (2009)
Cameroon	Areia, PB	NA	Polyculture	<i>Amyntas gracilis</i> , <i>Dichogaster</i> sp., <i>Pontoscolex corethrurus</i>	4	0/4	Guerra and Silva (1994)
	Eldorado, SP	Conventional	Monoculture	<i>Amyntas gracilis</i> , <i>Dichogaster</i> sp., <i>Pontoscolex corethrurus</i>	2	0/2	This study
	Jutaí River margin, AM	NA	NA	<i>Pontoscolex corethrurus</i>	ND	0/1	Righi (1990)
	Registro, SP	Conventional	Monoculture	<i>Amyntas gracilis</i> , <i>Pontoscolex corethrurus</i>	2	0/2	This study
	Sete Barras, SP	Conventional	Monoculture	<i>Amyntas gracilis</i> , <i>Pontoscolex corethrurus</i>	2	0/2	This study
	Mbalmayo Forest Reserve (low density cover)	Organic Agroforestry	Monoculture	<i>Dichogaster hauseri</i> , <i>Eminoscolex lamani</i> , <i>Eudrilidae</i> gen. et sp. nov.1 & 2, <i>Legonodrilus</i> sp. nov.1, <i>Malodrilus kamerunensis</i> , <i>Nematogenia panamaensis</i> , <i>Rosadrilus kamerunensis</i> , <i>Scolecillus tantillus</i>	8	7/1	Norgrove et al. (2011)
	Mbalmayo Forest Reserve (high density cover)	Organic Agroforestry	Monoculture	<i>Dichogaster annae</i> , <i>Dichogaster bolaii</i> , <i>Dichogaster</i> sp., <i>Eminoscolex lamani</i> , <i>Eudrilidae</i> sp., <i>Eudrilidae</i> gen. et sp. nov. 1, <i>Legonodrilus</i> sp. nov. 1, <i>Nematogenia panamaensis</i> , <i>Ocnerodrilidae</i> gen. et sp. nov., <i>Rosadrilus kamerunensis</i> , <i>Scolecillus tantillus</i>	10	7/3	Norgrove et al. (2011)
	Hainan Province	NA	NA	<i>Pheretima montana</i>	ND	0/1	Sun et al. (2012)
	Quindío, Circasia, Barcelona (La Sofe farm)	NA	Monoculture	<i>Aptodrilus fuhrmanni</i> , <i>Amyntas minimus</i> , <i>Glossodrilus chaguala</i> , <i>Glossodrilus panikita</i> , <i>Martiodrilus quimbayaensis</i>	5	4/1	Feijoo et al. (2018)
	Quindío, Circasia, Barcelona (La Sofe farm)	NA	Polyculture	<i>Aptodrilus fuhrmanni</i> , <i>Amyntas minimus</i> , <i>Glossodrilus chaguala</i> , <i>Glossodrilus panikita</i> , <i>Martiodrilus quimbayaensis</i>	5	4/1	Feijoo et al. (2018)
China	Quindío, Circasia, Barcelona (La Sofe farm)	NA	NA	<i>Amyntas gracilis</i> , <i>Perisoclex columbianus</i>	2	1/1	Feijoo et al. (2018)

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
Colombia	Armenia, Niagra (La Catalina)	NA	NA	<i>Amyntas gracilis</i> , <i>Glossodrilus griseus</i> , <i>Pontoscolex corethrurus</i>	3	1/2	Feijoo et al. (2018)
	Quindío, Calarcá, Quebrada Negra	NA	NA	<i>Glossodrilus griseus</i>	1	1/0	Feijoo et al. (2018)
	Quindío, Marmato (La Cristalina farm)	NA	Monoculture	<i>Glossodrilus lacteus</i>	1	1/0	Feijoo et al. (2018)
	Quindío, Marmato (La Cristalina farm)	NA	Polyculture	<i>Glossodrilus lacteus</i>	1	1/0	Feijoo et al. (2018)
	Quindío, Marmato (La Cristalina farm)	NA	NA	<i>Dichogaster affinis</i>	1	0/1	Feijoo et al. (2018)
	Armenia, La Revancha (Villa Sofia farm)	NA	NA	<i>Amyntas gracilis</i> , <i>Dichogaster affinis</i> , <i>Dichogaster bolau</i> , <i>Glossodrilus griseus</i> , <i>Perionyx excavatus</i>	5	1/4	Feijoo et al. (2018)
	Armenia, La Revancha (Bella Marina farm)	NA	NA	<i>Dichogaster saliens</i> , <i>Perisoclex columbianus</i>	2	1/1	Feijoo et al. (2018)
Costa Rica	Quindío, Armenia, El Rhin	NA	NA	<i>Perisoclex columbianus</i>	1	1/0	Feijoo et al. (2018)
	Quindío, Armenia, La India (La Ermita farm)	NA	NA	<i>Perisoclex coreguaje</i>	1	1/0	Feijoo et al. (2018)
	Circasia, Barcelona Baja rural (Buenos Aires farm)	NA	NA	<i>Amyntas gracilis</i> , <i>Dichogaster saliens</i> , <i>Pontoscolex corethrurus</i>	3	0/3	Feijoo et al. (2018)
	Quindío, Armenia, La India (La Miranda farm)	NA	NA	<i>Dichogaster saliens</i>	1	0/1	Feijoo et al. (2018)
	Quindío, Armenia, La Patria Cahuita	NA	NA	<i>Dichogaster saliens</i> <i>Pontoscolex corethrurus</i>	1 ND	0/1 ?/1	Feijoo et al. (2018) Lapied and Lavelle (2003)
Cuba	Boyeros	Organic	Monoculture	<i>Dichogaster affinis</i> , <i>Dichogaster bolau</i> , <i>Onychochaeta elegans</i> , <i>Polypheretima elongata</i> , <i>Protozapotecia angelesae</i>	5	2/3	Martínez-Leiva (2002)
Guadeloupe (France)	Capesterre-Belle-Eau	NA	Monoculture	<i>Pontoscolex corethrurus</i>	ND	?/1	Lafont et al. (2007)
	Capesterre-Belle-Eau (Gloria Bas)	Conventional	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
	Capesterre-Belle-Eau (Source)	Conventional	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
	Capesterre-Belle-Eau (Bergerie)	Agroecological	Monoculture	<i>Pontoscolex corethrurus</i> , unknown sp. 2	2	?	Burac et al. (2018)
	Baillif (Sextius)	Agroecological	Monoculture	<i>Pontoscolex corethrurus</i> , unknown sp.	2	?	Burac et al. (2018)
	Baillif (Grand Canon)	Agroecological	Monoculture	<i>Pontoscolex corethrurus</i> , unknown sp. 3	2	?	Burac et al. (2018)
	Saint-Claude (Saut d'Eau)	Agroecological	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
India	Dakshina Kannada District (Belthangady)	NA	NA	<i>Hoplochaetella kempi</i>	ND	1/0	Siddaraju et al. (2013)
	Dakshina Kannada District (Mangalore)	NA	NA	<i>Konkadrilus babli</i>	ND	1/0	Siddaraju et al. (2013)
	Dakshina Kannada District (Mangalore)	NA	NA	<i>Dichogaster affinis</i>	ND	0/1	Siddaraju et al. (2013)

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
India	Dakshina Kannada District (Bantwal)	NA	NA	<i>Octochaetona parva</i>	ND	1/0	Siddaraju et al. (2010)
	Dakshina Kannada District (sites not detailed)	NA	NA	<i>Amyntas corticis</i> , <i>Hoplochaetella kempi</i> , <i>Hoplochaetella stuarti</i> , <i>Hoplochaetella sutoria</i> , <i>Megascolex konkanensis</i> , <i>Metaphire posthuma</i> , <i>Octochaetona paliensis</i> , <i>Octochaetona parva</i>	ND	7/1	Siddaraju et al. (2010, 2013)
	Kerala (Vellayambalam)	NA	NA	<i>Perionyx excavatus</i> , <i>Pontoscolex corethrurus</i>	2	0/2	Nair et al. (2007)
	Mizoram	NA	Monoculture	<i>Drauidia nepalensis</i> , <i>Drauidia rangamaitiana</i> , <i>Drauidia</i> sp., <i>Metaphire houlleti</i> , <i>Perionyx excavatus</i>	5	3/2	Lalthanzara (2007)
	Mizoram	NA	Polyculture	<i>Drauidia nugana</i> , <i>Drauidia</i> sp., <i>Metaphire houlleti</i> , <i>Perionyx excavatus</i>	4	2/2	Lalthanzara (2007)
	Rajapalayam	NA	NA	<i>Lampito mauritii</i> , <i>Perionyx excavatus</i>	2	1/1	Mariappan et al. (2013)
	Udupi District (Adve)	NA	NA	<i>Megascolex konkanensis</i>	1	1/0	Kumar et al. (2018)
	Udupi District (Adve)	NA	NA	<i>Metaphire houlleti</i>	1	0/1	Kumar et al. (2018)
	Udupi District (Bellibetu)	NA	NA	<i>Metaphire houlleti</i> , <i>Pontoscolex corethrurus</i>	2	0/2	Kumar et al. (2018)
	Udupi District (Mudarangadi)	NA	NA	<i>Pontoscolex corethrurus</i>	1	0/1	Kumar et al. (2018)
	Udupi District (Nandikur)	NA	NA	<i>Drauidia ampullacea</i> , <i>Drauidia sulcata</i> , <i>Metaphire peguana</i>	3	3/0	Kumar et al. (2018)
	Udupi District (Nandikur)	NA	NA	<i>Drauidia ampullacea</i>	1	1/0	Kumar et al. (2018)
	Udupi District (Padabertu)	NA	NA	<i>Perionyx excavatus</i>	1	0/1	Kumar et al. (2018)
	Udupi District (Vellur)	NA	NA	<i>Mallebulla indica</i> , <i>Megascolex konkanensis</i>	2	2/0	Kumar et al. (2018)
	West Tripura (Mohanpur, Maheshkhola, Rastermatha)	Organic	Monoculture	<i>Amyntas alexandri</i> , <i>Drauidia assamensis</i> , <i>Drauidia papillifer</i> , <i>Eutyphoeus comillabhus</i> , <i>Lampito mauritii</i> , <i>Lennogaster</i> sp., <i>Metaphire houlleti</i> , <i>Metaphire posthuma</i> , <i>Octochaetona beatrix</i> , <i>Perionyx excavatus</i> , <i>Pontoscolex corethrurus</i>	3–7	4/7	Dhar and Chaudhuri (2018)
Indonesia	Bangkalan (Kamal, Bumeh, Socah, Bypass)	NA	NA	<i>Amyntas robustus</i> , <i>Metaphire californica</i> , <i>Metaphire javanica</i>	ND	1/2	Budijastuti (2019)
	Bangkalan (Tanah Merah)	NA	NA	<i>Metaphire posthuma</i>	1	0/1	Budijastuti (2019)
	Bangkalan (Labang)	NA	NA	<i>Amyntas robustus</i> , <i>Metaphire javanica</i> , <i>Metaphire californica</i> , <i>Pheretima racemosa</i>	4	2/2	Budijastuti (2019)
	Gresik (Driyorejo, Kedamean, Ngipik, SumengkoLegundi)	NA	NA	<i>Amyntas robustus</i> , <i>Metaphire javanica</i>	ND	1/1	Budijastuti (2019)
	Gresik (Wringinanamon)	NA	NA	<i>Amyntas robustus</i> , <i>Metaphire javanica</i> , <i>Metaphire posthuma</i>	3	1/2	Budijastuti (2019)
	Sidoarjo (Waru, Taman, Sidoarjo, Tulangan, Tanggulangin, Candi)	NA	NA	<i>Amyntas robustus</i> , <i>Metaphire javanica</i> , <i>Metaphire posthuma</i>	ND	1/2	Budijastuti (2019)
	Surabaya (Pakal, Benowo, Tandes, Sukolilo, Gubeng, Gununganyar)	NA	NA	<i>Amyntas robustus</i> , <i>Metaphire javanica</i> , <i>Metaphire posthuma</i>	ND	1/2	Budijastuti (2019)
	Lamto region	NA	NA	<i>Dichogaster wienkei</i> , <i>Reginaldia anomala</i> , <i>Stuhlmannia palustris</i> , <i>Stuhlmannia zizelae</i>	ND	4/0	Tondoh (1994)
Ivory Coast	Clarendon, Crofts Mountain	NA	NA	<i>Drauidia barwelli</i> , <i>Polypheretima elongata</i>	2	0/2	Sims (1987)
Jamaica	Ambatosoratra Ambatondrazaka	NA	NA	<i>Kynotus sihanakus</i> , <i>Kynotus</i> sp.2	2	2/0	Razafindrakoto et al. (2016), Csuzdi et al. (2017)
Madagascar							

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
Malaysia	Serdang, Sengalor (Universiti Putra Malaysia)	NA	NA	<i>Pontoscolex corethrurus</i>	ND	0/1	Teng et al. (2006)
	Le Lorrain (Feuillère)	Conventional	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
Martinique (France)	Le Lorrain (Limite)	Agroecological	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
	Le Lorrain (Bellevue)	Conventional	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
	L'Ajoupa-Bouillon (Allée Domergue 3)	Agroecological	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
	Basse-Pointe (Fromager Rivière)	Conventional	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
	Basse-Pointe (Dantu Bas)	Agroecological	Monoculture	<i>Pontoscolex corethrurus</i>	1	0/1	Burac et al. (2018)
Mexico	Tabasco	NA	Monoculture	<i>Lavellothrillus bonampakensis</i> , <i>Pontoscolex corethrurus</i>	2	1/1	Huerta et al. (2005)
	Tabasco, Teapa B1	NA	Monoculture	<i>Balanteodrilus pearsei</i> , <i>Drauidia barwelli</i> , <i>Polypheretima elongata</i> , <i>Pontoscolex corethrurus</i> , <i>Pontoscolex</i> sp.	5	1/4	Geissen et al. (2009)
	Tabasco, Teapa B2	NA	Monoculture	<i>Balanteodrilus pearsei</i> , <i>Dichogaster bolau</i> , <i>Drauidia barwelli</i> , <i>Periscolex brachycystis</i> , <i>Polypheretima elongata</i> , <i>Pontoscolex</i> sp.	6	2/4	Geissen et al. (2009)
	Tabasco, Teapa AF1	Agroforestry	Polyculture	<i>Balanteodrilus pearsei</i> , <i>Dichogaster bolau</i> , <i>Drauidia barwelli</i> , <i>Polypheretima elongata</i> , <i>Pontoscolex corethrurus</i> , <i>Pontoscolex</i> sp.	6	2/4	Geissen et al. (2009)
	Tabasco, Teapa AF2	Agroforestry	Polyculture	<i>Balanteodrilus pearsei</i> , <i>Dichogaster bolau</i> , <i>Drauidia barwelli</i> , <i>Polypheretima elongata</i> , <i>Pontoscolex corethrurus</i>	5	1/4	Geissen et al. (2009)
	Tabasco, Teapa (site 1)	Conventional	NA	<i>Dichogaster saliens</i> , <i>Diplolema murchiei</i> , <i>Pontoscolex corethrurus</i>	3	1/2	Huerta et al. (2013)
	Tabasco, Teapa (site 2)	Conventional	NA	<i>Dichogaster saliens</i> , <i>Pontoscolex corethrurus</i>	2	0/2	Huerta et al. (2013)
	Tabasco, Teapa (site 3)	Conventional	NA	<i>Diplolema murchiei</i> , <i>Polypheretima elongata</i> , <i>Pontoscolex corethrurus</i>	3	1/2	Huerta et al. (2013)
	Tabasco, Teapa (site 4)	Conventional	Polyculture	<i>Amyntas gracilis</i> , <i>Pontoscolex corethrurus</i>	2	0/2	Huerta et al. (2007)
	Tabasco, Teapa (site 5)	Conventional	Polyculture	<i>Dichogaster saliens</i> , <i>Polypheretima elongata</i> , <i>Pontoscolex corethrurus</i>	3	0/3	Huerta et al. (2013)
	Tabasco, Teapa (site 6)	Conventional	NA	<i>Dichogaster saliens</i> , <i>Pontoscolex corethrurus</i>	2	0/2	Huerta et al. (2007)
	Tabasco, Pablo L. Sidar	NA	Monoculture	<i>Lavellothrillus bonampakensis</i> , <i>Pontoscolex corethrurus</i>	2	1/1	Huerta et al. (2013)
Nicaragua	Tamaulipas (Biosphere Reserve “El Cielo”)	NA	NA	<i>Amyntas gracilis</i>	ND	0/1	Barois (1992)
	Actopan, Ejido Buenavista	NA	NA	<i>Balanteodrilus psammophilus</i>	ND	1/0	Fragoso and Rojas (2007)
Peru	Managua	NA	NA	<i>Dichogaster bolau</i> , <i>Periscolex brachycystis</i>	2	1/1	Sherlock et al. (2011)
	Sarita Colonia	NA	Monoculture	<i>Pontoscolex corethrurus</i> and two native spp.	3	2/1	Pashanasi (2007)
Philippines	Davao (Sumitomo Fruits Corporation, 15% site)	Conventional	Monoculture	<i>Metaphire</i> sp., <i>Pithemera bicincta</i> , <i>Pontoscolex corethrurus</i>	3	1/2	Fusilero et al. (2013)
	Davao (Sumitomo Fruits Corporation, 25% site)	Conventional	Monoculture	<i>Metaphire cai</i> , <i>Metapheretima</i> sp., <i>Perionyx excavatus</i>	3	2/1	Fusilero et al. (2013)
Portugal	Madeira Island (Ribeira Brava)	NA	NA	<i>Aporrectodea moebii</i> , <i>Eisenia eisens</i> , <i>Metaphire californica</i>	3	0/3	Talavera (1996)
	Madeira Island (Funchal)	NA	NA	<i>Amyntas gracilis</i> , <i>Metaphire californica</i> , <i>Ocnodrilus occidentalis</i>	3	0/3	Talavera (1996)

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
Portugal	Madeira Island (Santa Cruz)	NA	NA	<i>Anyynthas gracilis</i>	1	0/1	Talavera (1996)
	Madeira Island (Terceira Lombada)	NA	NA	<i>Aporrectodea moebii</i> , <i>Eiseniella tetraedra</i>	2	0/2	Talavera (1996)
	Madeira Island (Porto Moniz)	NA	NA	<i>Anyynthas gracilis</i> , <i>Aporrectodea rosea</i> , <i>Aporrectodea trapezoides</i> , <i>Dendrobaena pseudohortensis</i>	4	0/4	Talavera (2011)
Seychelles	Madeira Island (Terceira Lombada)	NA	NA	<i>Aporrectodea caliginosa</i> , <i>Aporrectodea rosea</i> , <i>Eiseniella tetraedra</i>	3	0/3	Talavera (2011)
	Cousine Island	NA	Monoculture	<i>Pontoscolex corethrurus</i>	ND	0/1	Plisko (2001)
	KwaZulu-Natal (Fairfield Farm)	NA	Monoculture	<i>Pontoscolex corethrurus</i>	ND	0/1	Plisko (2001)
South Africa	KwaZulu-Natal (Benhurst Farm)	NA	Monoculture	<i>Pontoscolex corethrurus</i>	ND	0/1	Plisko (2001)
	KwaZulu-Natal (6 sites in Eshowe)	NA	Monoculture	<i>Anyynthas corticis</i> , <i>Anyynthas minimus</i> , <i>Anyynthas rodericensis</i> , <i>Dichogaster bolau</i> , <i>Pontoscolex corethrurus</i> , and one other sp.	ND	0/5	Dlamini and Haynes (2004)
Spain	Gomera Island (Agulo)	NA	NA	<i>Anyynthas rodericensis</i> , <i>Allolobophora chlorotica</i> , <i>Eiseniella tetraedra</i> , <i>Ocnoderilus occidentalis</i>	4	0/4	Talavera (1990a, 2007)
	Gomera Island (Barranco de la Villa)	NA	NA	<i>Bimastos rubidus</i> , <i>Ocnoderilus occidentalis</i> , <i>Pithemera bicincta</i>	3	0/3	Talavera (2007)
	Gomera Island (Barranco del Valle)	NA	NA	<i>Allolobophora chlorotica</i> , <i>Metaphire californica</i>	2	0/2	Talavera (1990b, 2007)
	Gomera Island (Casas de Aluce)	NA	NA	<i>Aporrectodea rosea</i> , <i>Microscolex phosphoreus</i>	2	0/2	Talavera (2007)
	Gomera Island (Cabo Verde)	NA	NA	<i>Anyynthas gracilis</i> , <i>Bimastos rubidus</i>	2	0/2	Talavera (1990b, 2007)
	Gomera Island (Costa Agulo)	NA	NA	<i>Aporrectodea trapezoides</i> , <i>Anyynthas rodericensis</i> , <i>Bimastos rubidus</i> , <i>Ocnoderilus occidentalis</i>	4	0/4	Talavera (2007)
	Gomera Island (El Molinito)	NA	NA	<i>Anyynthas morrisi</i> , <i>Microscolex phosphoreus</i>	2	0/2	Talavera (2007)
	Gomera Island (Hermigua)	NA	NA	<i>Aporrectodea rosea</i> , <i>Bimastos rubidus</i> , <i>Eisenia fetida</i> , <i>Ocnoderilus occidentalis</i>	4	0/4	Talavera (1990a, 2007)
	Gomera Island (Laguna de Santiago)	NA	NA	<i>Anyynthas morrisi</i> , <i>Aporrectodea rosea</i> , <i>Aporrectodea trapezoides</i> , <i>Bimastos rubidus</i> , <i>Dendrobaena hortensis</i> , <i>Dichogaster affinis</i> , <i>Metaphire californica</i> , <i>Pithemera bicincta</i>	9	0/9	Talavera (2007)
	Gomera Island (Playa de Santiago)	NA	NA	<i>Ocnoderilus occidentalis</i>	1	0/1	Talavera (1990a)
	Gomera Island (Seimal)	NA	NA	<i>Eiseniella tetraedra</i> , <i>Metaphire californica</i> , <i>Microscolex phosphoreus</i>	3	0/3	Talavera (2007)
	Gomera Island (Taguluche)	NA	NA	<i>Anyynthas morrisi</i> , <i>Allolobophora chlorotica</i> , <i>Octalasion lacteum</i>	3	0/3	Talavera (2007)
	Gomera Island (Valle Gran Rey)	NA	NA	<i>Allolobophora chlorotica</i> , <i>Aporrectodea trapezoides</i> , <i>Dendrobaena hortensis</i> , <i>Eisenia fetida</i> , <i>Microscolex dubius</i> , <i>Pithemera bicincta</i>	5	0/5	Talavera (2007)
	Gran Canaria (Lomo del Galeón)	NA	NA	<i>Ocnoderilus occidentalis</i>	1	0/1	Talavera (1990a)
	Gran Canaria (Los Llanos)	NA	NA	<i>Ocnoderilus occidentalis</i> , <i>Pithemera bicincta</i>	2	0/2	Talavera (1990a)

Gran Canaria Island (Bañaderos)	NA	NA	NA	<i>Metaphire californica</i>	1	0/1	Talavera (1990b)
Gran Canaria Island (Barranco Guinguada)	NA	NA	NA	<i>Anyynthas morrisi</i>	1	0/1	Talavera (1990b)
Gran Canaria Island (Frontón)	NA	NA	NA	<i>Anyynthas gracilis</i>	1	0/1	Talavera (1990b)
Gran Canaria Island (Galdar)	NA	NA	NA	<i>Anyynthas morrisi</i>	1	0/1	Talavera (1990b)

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
Spain	Gran Canaria Island (Hoya Mondondo)	NA	NA	<i>Pithemera bicincta</i>	1	0/1	Talavera (1990b)
	Gran Canaria Island (La Aldea)	NA	NA	<i>Dichogaster affinis</i>	1	0/1	Talavera (1992b)
	Gran Canaria Island (Pedrazo)	NA	NA	<i>Pithemera bicincta</i>	1	0/1	Talavera (1990b)
	Gran Canaria Island (Tenoya)	NA	NA	<i>Amyntas morrisi</i>	1	0/1	Talavera (1990b)
	Hierro Island (Los Mocanes)	NA	NA	<i>Ocnodrilus occidentalis</i>	1	0/1	Talavera (1990a)
	Hierra Island (NE tip)	NA	NA	<i>Microscolex phosphoreus</i>	ND	0/1	Talavera and Pérez (2009)
	La Palma Island (Barranco de las Angustias)	NA	NA	<i>Amyntas gracilis</i>	1	0/1	Talavera (1990b)
	La Palma Island (Barranco Nogales)	NA	NA	<i>Amyntas gracilis</i>	1	0/1	Talavera (1990b)
	La Palma Island (El Socorro)	NA	NA	<i>Pithemera bicincta</i>	1	0/1	Talavera (1990b)
	La Palma Island (La Caldereta)	NA	NA	<i>Amyntas morrisi, Metaphire californica</i>	2	0/2	Talavera (1990b)
	La Palma Island (Los Cancajos)	NA	NA	<i>Amyntas morrisi</i>	1	0/1	Talavera (1990b)
	La Palma Island (Los Llanos de Aridane)	NA	NA	<i>Amyntas morrisi, Metaphire californica</i>	2	0/2	Talavera (1990b)
	La Palma Island (Tazacorte)	NA	NA	<i>Amyntas gracilis, Amyntas morrisi, Metaphire californica</i>	3	0/3	Talavera (1990b)
	Tenerife Island (Abama)	NA	Monoculture	<i>Aporrectodea rosea, Dendrobaena hortensis, Eisenia andrei, Microscolex dubius</i>	4	0/4	Talavera (1992a)
	Tenerife Island (Adeje)	NA	NA	<i>Ocnodrilus occidentalis</i>	1	0/1	Talavera (1990a)
	Tenerife Island (Bajamar)	NA	Monoculture	<i>Amyntas morrisi, Aporrectodea rosea, Dichogaster affinis, Eisenia andrei, Microscolex phosphoreus, Ocnodrilus occidentalis</i>	6	0/6	Talavera (1990a, 1992a, 1992b)
	Tenerife Island (Barranco de Santos)	NA	Monoculture	<i>Amyntas morrisi, Aporrectodea rosea, Bimastos rubidus, Eisenia andrei, Microscolex dubius, Pithemera bicincta</i>	6	0/6	Talavera (1990b, 1992a)
	Tenerife Island (Barranco del Inglés)	NA	Monoculture	<i>Aporrectodea rosea, Aporrectodea trapezoides, Eisenia andrei, Microscolex dubius</i>	4	0/4	Talavera (1992a)
	Tenerife Island (Barranco la Atalaya)	NA	Monoculture	<i>Aporrectodea rosea, Pithemera bicincta</i>	2	0/2	Talavera (1992a)
	Tenerife Island (Barranco las Galletas)	NA	Monoculture	<i>Aporrectodea rosea, Eisenia andrei, Ocnodrilus occidentalis</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Barranco San Felipe)	NA	Monoculture	<i>Amyntas gracilis, Eisenia andrei, Pithemera bicincta</i>	3	0/3	Talavera (1992a, 1990b)
	Tenerife Island (Buenavista del Norte)	NA	NA	<i>Ocnodrilus occidentalis</i>	1	0/1	Talavera (1990a)
	Tenerife Island (Casablanca)	NA	Monoculture	<i>Amyntas corticis, Aporrectodea rosea, Eisenia andrei, Ocnodrilus occidentalis</i>	4	0/4	Talavera (1992a)
	Tenerife Island (Costa Valle Guerra)	NA	Monoculture	<i>Amyntas gracilis</i>	1	0/1	Talavera (1992a)
	Tenerife Island (El Puente)	NA	Monoculture	<i>Amyntas gracilis, Aporrectodea rosea, Eisenia andrei, Microscolex phosphoreus, Ocnodrilus occidentalis</i>	5	0/5	Talavera (1992a)
	Tenerife Island (El Rincón)	NA	Monoculture	<i>Amyntas gracilis, Bimastos rubidus, Dendrobaena cognetti, Microscolex dubius, Microscolex phosphoreus, Octodrilus complanatus</i>	6	0/6	Talavera (1992a)

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
Spain	Tenerife Island (El Socorro)	NA	NA	<i>Pithemera bicincta</i>	1	0/1	Talavera (1990b)
	Tenerife Island (Fanabé)	NA	Monoculture	<i>Anyinthus corticis</i> , <i>Aporrectodea rosea</i> , <i>Dichogaster affinis</i> , <i>Eisenia andrei</i> , <i>Ocnodrilus occidentalis</i>	5	0/5	Talavera (1990a, 1992a, 1992b)
	Tenerife Island (Güimar)	NA	NA	<i>Dichogaster affinis</i> , <i>Ocnodrilus occidentalis</i>	2	0/2	Talavera (1990a, 1992b)
	Tenerife Island (Iboybo)	NA	Monoculture	<i>Aporrectodea rosea</i> , <i>Eisenia andrei</i> , <i>Ocnodrilus occidentalis</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Icod de Los Vinos)	NA	Monoculture	<i>Dendrobaena cognetti</i> , <i>Bimastos rubidus</i> , <i>Octodrilus complanatus</i> , <i>Ocnodrilus occidentalis</i>	4	0/4	Talavera (1992a)
	Tenerife Island (Igueste)	NA	Monoculture	<i>Allolobophora chlorotica</i> , <i>Aporrectodea rosea</i> , <i>Aporrectodea trapezoides</i> , <i>Pontoscolex corethrurus</i> , <i>Ocnodrilus occidentalis</i>	5	0/5	Talavera (1992a)
	Tenerife Island (La Hondura)	NA	Monoculture	<i>Anyinthus morrisi</i>	1	0/1	Talavera (1992a)
	Tenerife Island (La Longuera)	NA	Monoculture	<i>Anyinthus morrisi</i> , <i>Aporrectodea rosea</i> , <i>Eisenia fetida</i> , <i>Microscolex dubius</i> , <i>Octodrilus complanatus</i>	5	0/5	Talavera (1992a)
	Tenerife Island (La Matanza)	NA	Monoculture	<i>Bimastos rubidus</i> , <i>Eisenia andrei</i> , <i>Microscolex phosphoreus</i>	3	0/3	Talavera (1992a)
	Tenerife Island (La Montañera)	NA	NA	<i>Pithemera bicincta</i>	1	0/1	Talavera (1990b)
	Tenerife Island (La Vera)	NA	Monoculture	<i>Bimastos rubidus</i> , <i>Eisenia andrei</i> , <i>Microscolex phosphoreus</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Las Arenas)	NA	NA	<i>Anyinthus morrisi</i>	1	0/1	Talavera (1990b)
	Tenerife Island (Las Galletas)	NA	Monoculture	<i>Eisenia andrei</i> , <i>Bimastos eiseni</i> , <i>Ocnodrilus occidentalis</i>	3	0/3	Talavera (1990a, 1992a)
	Tenerife Island (Las Madrigueras)	NA	NA	<i>Anyinthus morrisi</i>	1	0/1	Talavera (1990b)
	Tenerife Island (Los Quintos)	NA	Monoculture	<i>Dendrobaena cognetti</i> , <i>Bimastos rubidus</i> , <i>Microscolex phosphoreus</i> , <i>Pithemera bicincta</i> , <i>Ocnodrilus occidentalis</i>	5	0/5	Talavera (1992a)
	Tenerife Island (Los Realejos)	NA	NA	<i>Pithemera bicincta</i>	1	0/1	Talavera (1990b)
	Tenerife Island (Los Rechazos)	NA	Monoculture	<i>Aporrectodea trapezoides</i> , <i>Bimastos rubidus</i> , <i>Eisenia fetida</i> , <i>Octodrilus complanatus</i> , <i>Pithemera bicincta</i>	5	0/5	Talavera (1992a)
	Tenerife Island (Los Silos)	NA	Monoculture	<i>Anyinthus morrisi</i> , <i>Aporrectodea rosea</i> , <i>Dichogaster affinis</i> , <i>Eisenia andrei</i> , <i>Ocnodrilus occidentalis</i>	5	0/5	Talavera (1992a, 1992b)
	Tenerife Island (Loss Llanos)	NA	Monoculture	<i>Anyinthus morrisi</i> , <i>Bimastos rubidus</i> , <i>Eisenia andrei</i> , <i>Pithemera bicincta</i>	4	0/4	Talavera (1992a)
	Tenerife Island (Playa de las Aguas)	NA	Monoculture	<i>Anyinthus morrisi</i> , <i>Eisenia andrei</i> , <i>Pithemera bicincta</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Playa de San Juan)	NA	Monoculture	<i>Aporrectodea rosea</i> , <i>Dendrobaena hortensis</i> , <i>Bimastos rubidus</i> , <i>Eisenia andrei</i>	4	0/4	Talavera (1992a)
	Tenerife Island (Playa San Marcos)	NA	Monoculture	<i>Pithemera bicincta</i> , <i>Bimastos rubidus</i> , <i>Microscolex phosphoreus</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Puertito de Gíllimar)	NA	Monoculture	<i>Microscolex phosphoreus</i> , <i>Pithemera bicincta</i> , <i>Ocnodrilus occidentalis</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Puerto de Santiago)	NA	Monoculture	<i>Anyinthus morrisi</i>	1	0/1	Talavera (1990b, 1992a)
	Tenerife Island (Punta del Hidalgo)	NA	Monoculture	<i>Anyinthus gracilis</i> , <i>Ocnodrilus occidentalis</i>	2	0/2	Talavera (1992a)
	Tenerife Island (San Andrés)	NA	Monoculture	<i>Anyinthus morrisi</i> , <i>Aporrectodea rosea</i> , <i>Microscolex phosphoreus</i> , <i>Ocnodrilus occidentalis</i>	4	0/4	Talavera (1992a)
	Tenerife Island (San Bernardo)	NA	Monoculture	<i>Anyinthus corticis</i> , <i>Anyinthus morrisi</i> , <i>Aporrectodea rosea</i> , <i>Eisenia andrei</i>	4	0/4	Talavera (1992a)

Country	Location	Management	Culture type	Earthworm species	Richness	Native (N) / Exotic (E)	References
Spain	Tenerife Island (San Juan de la Rambla)	NA	Monoculture	<i>Amyntas gracilis</i> , <i>Bimastos rubidus</i> , <i>Dendrobaena hortensis</i> , <i>Eisenia fetida</i> , <i>Pithechera bicincta</i>	5	0/5	Talavera (1990b, 1992a)
	Tenerife Island (San Pedro de Daure)	NA	Monoculture	<i>Amyntas morrisi</i> , <i>Aporrectodea rosea</i> , <i>Ocnodrilus occidentalis</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Santo Domingo)	NA	Monoculture	<i>Dendrobaena cognetti</i> , <i>Microscolex dubius</i> , <i>Microscolex phosphoreus</i>	3	0/3	Talavera (1992a)
	Tenerife Island (Taganana)	NA	Monoculture	<i>Amyntas morrisi</i>	1	0/1	Talavera (1992a)
	Tenerife Island (Tejina)	NA	Monoculture	<i>Amyntas corticis</i> , <i>Pithechera bicincta</i>	2	0/2	Talavera (1992a)
Taiwan	Central region	NA	NA	<i>Pontoscolex corethrurus</i>	ND	0/1	Tsai et al. (2000)
Uganda	Kabanyolo University Farm	NA	NA	<i>Dichogaster</i> sp. 1, <i>Dichogaster</i> sp. 2, <i>Gordiodrilus</i> sp., <i>Pygmaeodrilus</i> sp., <i>Polytoreutus</i> sp. 1	5	5/0	Block and Banage (1968)

Table 5. Number of quantitative (with abundance data) and qualitative (where species were identified) sampling sites and earthworm species (total, native, and exotic) and families found in banana plantations in different countries of the world.

Country	No. sites: Quant./Qual. ¹	Total No. species	Native	Exotic	Families
Asia	6/≥47	35	22	13	5
Bangladesh	0/1	3	1	2	2
China	0/1	1	0	1	1
India	4/≥20	27	20	7	5
Indonesia	0/23	5	1	4	1
Malaysia	0/1	1	0	1	1
Philippines	2/2	6	3	3	2
Taiwan	0/1	1	0	1	1
Africa	33/97	50	20	30	7
Cameroon	10/2	12	9	3	3
Canary Islands (Spain) ²					
<i>Gomera</i>	0/13	18	0	18	4
<i>Gran Canaria</i>	0/10	6	0	6	3
<i>Hierro</i>	0/2	2	0	2	2
<i>La Palma</i>	0/7	3	0	3	1
<i>Tenerife</i>	0/45	19	0	19	5
Ivory Coast	1/1	4	4	0	3
Madagascar	0/1	2	2	0	1
Madeira (Portugal) ²	0/6	10	0	10	3
Seychelles	0/1	1	0	1	1
South Africa	6/8	5	0	5	3
Uganda	6/1	5	5	0	3
North America	12/16	14	7	7	4
Bermuda	0/2	2	0	2	1
Mexico	12/14	12	5	7	4
Central America/Caribbean	53/≥17	≥10	4	≥6	4
Costa Rica	≥5/1	1	0	1	1
Cuba	1/1	5	2	3	3
Dominica	1/0	2	1?	1	2
Guadeloupe (France)	40/7	4?	?	≥1	≥1
Martinique (France)	6/6	1	0	1	1
Jamaica	0/1	2	0	2	2
Nicaragua	0/1	2	1	1	2
South America	49/33	20	10	10	6
Brazil	16/16	7	0	7	5
Colombia	32/15	15	8	7	4
Peru	1/1	3	2	1	≥1
Total	153/210	≥104	≥61	≥43	10

¹Quant.=quantitative samples, taken using various sampling methods (mostly hand sorting of soil monoliths); Qual.=qualitative samples, usually performed for biodiversity studies (species presence) and normally without specifying volume of soil sampled; ²Although politically these islands belong to Europe, biogeographically they belong to Africa.

Table 4), the role of less intensive banana production systems in maintaining native earthworm populations must still be further evaluated.

High species richness was also detected overall in Spain (25), mainly due to the higher sampling effort involving a large number of sites in the Canary Islands. However, all of the species encountered on the islands offshore of Africa were exotic, their introduction having been stimulated over centuries of human colonization bringing in exotic soils and crops (Talavera 2007, 2011). The Caribbean islands had few species (5),

despite a large sampling effort, and many sites were dominated by *P. corethrurus* (Burac et al. 2018). In Brazil, Costa Rica, Martinique, Jamaica, Bermuda, the Seychelles, Taiwan, Malaysia, and China, all the earthworm species encountered were exotic (Table 5). The continent with the highest number of species recorded was Africa (50), of which 40% were native. In Asia, 35 species were recorded, with a higher proportion of natives (66%). In North and South America, around 50% of the species found were native, but these were mainly due to the higher number of natives observed in Colombian (Feijoo et al. 2018) and Mexican (Geissen et al. 2009; Huerta et al. 2013) plantations.

Species richness in individual banana/plantain fields was measured in 166 of the 210 sites, and was generally very low, with an overall mean of 2.7 species per site worldwide, of which less than one (0.5) was native and 2.1 were exotic (full dataset in Cremonesi et al. 2020). Absolute richness in an individual plantation was highest in the banana plantations in Cameroon (Norgrove et al. 2011), where 8 and 10 species were found (Table 4), most of them native. The only other place with such high richness was a plantation in Gomera Island (Laguna de Santiago), where 9 species were found (Talavera 2007), although all of them were exotic. In West Tripura, up to 7 species were found in a banana plantation (Dhar and Chaudhuri 2018), but most plantations in the world had less than 3 species (~70% of sites), and the highest proportion was of sites with only 1 species (~30% of sites).

There was a clear positive relationship between the number of sites sampled in each country and the total number of species encountered ($r = 0.7$, $p < 0.01$), particularly for exotic ($r = 0.78$, $p < 0.01$) species (Fig. 3A). Although also positive, this relationship was not significant for native species. Nonetheless, the species accumulation curve for native species for all sampling sites in the world revealed a steep slope, that contrasts with the flattened-out accumulation curves for total and exotic species (Fig. 3B). This indicates that greater sampling efforts, particularly in more low-input production systems, especially in tropical countries with high earthworm biodiversity such as Ecuador (no studies with earthworms identified yet), Brazil and Colombia (Brown and James 2007; Feijoo 2007; Zicsi 2007) will certainly increase the number of species known from banana/plantain fields. Greater sampling efforts are also needed in other tropical countries with important plantain/banana production (FAO 2018), particularly when intercropped or in agroforestry systems (Norgrove et al. 2011; Norgrove and Hauser 2014), and where mostly native earthworm species may inhabit these fields, such as seen for Cameroon, Uganda and Ivory Coast. This phenomenon may likely also be applicable to other Western, Central and Eastern African countries, as well as many other Asian and Pacific countries, but the paucity of available data impedes further speculation.

Of the over 100 species found in banana and plantain fields worldwide, most belonged to the Megascolecidae (22%), Lumbricidae (17%) and Acanthodrilidae (16%) families (Cremonesi et al. 2020). These widespread exotic and often invasive species are found throughout the tropics and subtropics, and include several *Amyntas* and *Metaphire* spp. (Blakemore 2002). The most consistently recorded megascolecids were *A. gracilis* (6% of all records), *Amyntas morrisi* (Beddard, 1892) (5%), *Pithemera bicincta* (Perrier, 1875) (4%) and *Metaphire californica* (Kingerg, 1867), *Perionyx*

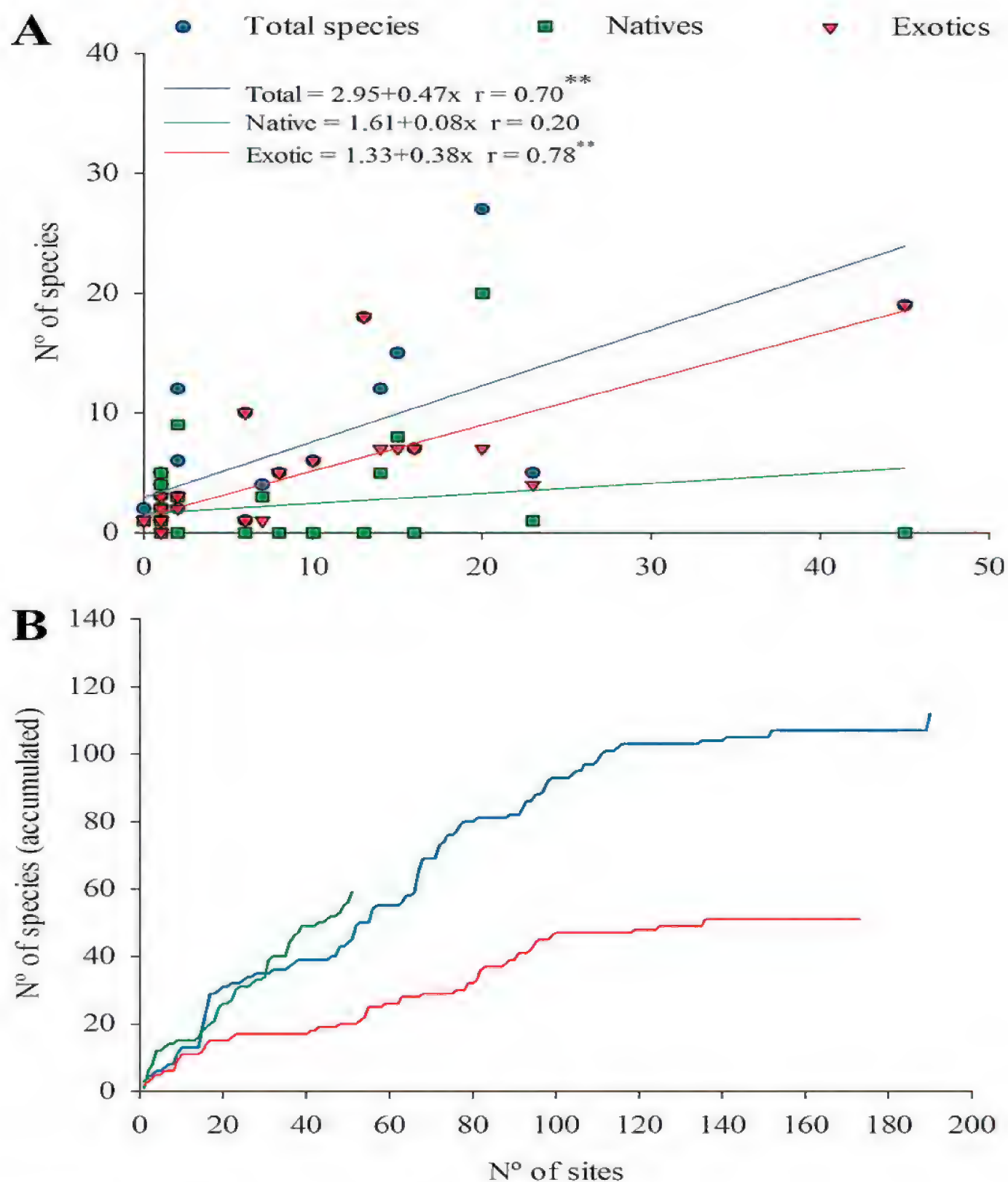


Figure 3. A Relationship between species richness (total, native, and exotic species) and the number of sampling sites in each world country (data from Table 5) and **B** Species accumulation curves for total, native and exotic species, depending on the number of sampling sites across the world. Linear regression equations and the value and significance (p value, with ** indicating $p < 0.01$) of the Pearson correlation coefficient (r) are provided in (**A**).

excavatus Perrier, 1872 and *Polypheretima elongata* (Perrier, 1872) (all with 2% each) (Cremonesi et al. 2020). These megascolecids were found in over 15 countries, and were especially frequent in the Canary Islands. All of the lumbricids reported were exotic, and mainly found in the Canary and Madeira Islands (Spain, Portugal), with *Aporrectodea rosea* (Savigny, 1826) and *Eisenia andrei* Bouché, 1972 (both with ~4%)

and *Bimastos rubidus* (Savigny, 1826) (3%) being the most frequently reported. Various octochaetid *Dichogaster* spp. of the Benhamiinae subfamily, i.e., *Dichogaster* (*Diplothecondrilus*) *affinis* (Michaelsen, 1890), *D. (D.) bolau* (Michaelsen, 1891) and *D. (D.) saliens* (Beddard, 1893) (all with around 2% each) and the acanthodrilinae *Microscolex* spp., i.e., *Microscolex phosphoreus* (Dugés, 1837) with 3% and *M. dubius* (Fletcher, 1887) with 2% of records, were the most reported acanthodrilids. The *Dichogaster* spp. were found in 11 countries, mainly in Latin America and the Canary Islands, while the *Microscolex* spp. were found only in the Canary Island banana plantations. Similarly, the ocnerodrilid *Ocnerodrilus occidentalis* Eisen, 1878 with 6% of all records, was found in three countries (Brazil, Portugal, Spain), but most frequently in the Canary Islands.

The most commonly encountered earthworm species in banana plantations was *P. corethrurus* (11%), found in 15 countries, mainly in Latin America, but also in places as far away as South Africa, India, Bangladesh, Malaysia, Philippines and Taiwan. Interestingly, 37 out of 54 sites (69%) that identified earthworm species reported *P. corethrurus* as dominant in the banana plantations (Table 3).

Although *P. corethrurus* may affect soil physical properties negatively by increasing soil compaction under some conditions, it can also positively affect biogeochemical processes, microbial activity, plant production, and soil recovery (see review in Taheri et al. 2018). Furthermore, this species is known to reduce plant-parasitic nematode incidence in banana plants (Loranger-Merciris et al. 2012), and has also been known to promote beneficial plant growth-promoting bacteria in the rhizosphere (Braga et al. 2015). Hence, further work is warranted on the potential beneficial impacts of the presence and populations of *P. corethrurus* on banana plants, particularly considering its widespread distribution and high abundance in some locations (e.g., Costa Rica, Brazil, Guadeloupe, Martinique, Mexico). Several megascolecs such as *A. gracilis* are also known to affect soil physical and chemical properties in annual cropping systems (e.g., Peixoto and Marochi 1996; Bartz et al. 2010) as well as crop production (Brown et al. 1999), but little is known of their effects on banana plants. The latter statement is also valid for all of the other species most commonly found in banana plantations.

A total of 31 studies performed in 153 sites and 15 countries (Table 5) had quantitative earthworm data (on abundance and/or biomass) taken mainly by hand sorting soil monoliths of variable size (mostly 25 x 25 cm but sometimes larger, e.g., 50 x 50 cm) and occasionally using liquid extraction (e.g., formalin expulsion). Most of the study sites were in Guadeloupe ($N = 40$, of which 34 were by Clermont-Dauphin et al. (2004) and Colombia ($N = 32$; Molina and Feijoo 2017).

Overall earthworm abundance ranged from a minimum of 0 (Figuerola 2019) in an Ecuadorian plantation, to a maximum mean of over 1500 indiv. m⁻² in banana plantations in Kwazulu-Natal, South Africa (Dlamini and Haynes 2004). Maximum biomass attained was 453.6 g m⁻² for a site in West Tripura, India (Dhar and Chaudhuri 2018). Interestingly, a large number of sites (>50) had abundance values over 100 indiv. m⁻², which could be considered quite high for earthworm density in annual agricultural crops (Bartz et al. 2013). Nonetheless, bananas are perennials often cultivated over

several cropping cycles, allowing for reduced negative effects of soil preparation, and the soils are also often limed to correct pH and fertilized with inorganic fertilizers (mainly N, P and K) to promote soil fertility and banana production. In these conditions, earthworms present find a soil protected from rainfall impact, as well as frequent organic matter additions through the management of the banana trees, particularly where the residues are left on the soil surface. Consequently, their populations can increase rather rapidly over time, as observed by Okwakol (1994) in Uganda (Table 3).

These high earthworm abundances and biomasses may be contributing significantly to soil processes (bioturbation, nutrient cycling) in banana/plantain fields, as biomasses over 17 g m^{-2} and above 32 g m^{-2} are known to lead to moderate (20–40%) and important (>40%) grain production increases, respectively (Brown et al. 1999). Earthworm-induced improvement of plant health and production includes, e.g., plant-parasitic nematode population control (Lafont et al. 2007; Loranger-Merciris et al. 2012), high stable bioaggregate formation, creation of many galleries in the soil and enhanced nutrient mineralization (Lavelle 1997), all factors that deserve future attention. On the other hand, low earthworm abundance may be an indicator of soil degradation, or the use of inappropriate management practices, such as soil inversion or toxic pesticide use (Demetrio et al. 2019). This type of information could be used to help farmers with their management decisions, such as reduction in nematicide applications that reduce earthworm populations (Clermont-Dauphin et al. 2004).

Finally, 18 of the major banana-producing countries in the world (34 countries with >30,000 ha in production, or >1 Million T bananas produced yr^{-1} ; FAO 2018) were not examined in the present review due to lack of data. Hence, further sampling efforts are needed in order to provide adequate information on earthworm abundance and biodiversity in banana plantations in these countries, and to complement those reported here but with low sample intensity, particularly focusing on the presence of native species and/or large earthworm abundances, and to identify the reasons for these phenomena and their consequences for banana production and biodiversity conservation.

Conclusions

Earthworms are an important component of banana and plantain fields worldwide and deserve further attention by taxonomists, ecologists and agronomists. Under some conditions, especially in lower-input polycultures, their abundance and biomass may reach high values and contribute significantly to soil processes and plant production. More than 70 studies performed in over 200 banana plantations of 28 countries found >100 species (around 60% of them native) from 10 families, although species richness in each sited tended to be low (generally <3 species) and exotic species predominated (particularly *P. corethrurus*). However, as many important banana-producing countries have not yet been evaluated, further work is warranted in order to better understand the earthworm communities and their functional roles in plantain/banana fields, and the role of management practices in affecting their populations and diversity worldwide.

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References

- ABAVAR – Associação dos Bananicultores do Vale do Ribeira (2015) Dia Mundial da Banana. ABAVAR, São Paulo. <http://abavar.com.br/Noticias/2015/09/DiaMundialDaBanana.pdf>
- Agüero R, Rojas S, Pérez L (2002) Poblaciones de lombrices bajo seis estrategias de manejo de malezas en una plantación de banano. *Agronomía Mesoamericana* 13(1): 25–29. <https://doi.org/10.15517/am.v13i1.13234>
- Anderson JM, Ingram JSI (1993) Tropical soil biology and fertility: a handbook of methods (2nd Edn.). CAB International, Wallingford, 171 pp.
- Avilés DFV (2017) Biodiversidad intraespecífica varietal para mejorar ambientes degradados por monocultivos en Musáceas, como medida de control de plagas y enfermedades. PhD thesis, Universitat Autònoma de Barcelona, Spain.
- Baretta D, Santos JCP, Segat JC, Geremia EV, Oliveira Filho LD, Alves MV (2011) Fauna edáfica e qualidade do solo. *Tópicos em Ciência do Solo* 7: 119–170.
- Barois I (1992) Mucus production and microbial activity in the gut of two species of *Amyntas* (Megascolecidae) from cold and warm tropical climates. *Soil Biology and Biochemistry* 24(12): 1507–1510. [https://doi.org/10.1016/0038-0717\(92\)90141-J](https://doi.org/10.1016/0038-0717(92)90141-J)
- Barois I, Cadet P, Albrecht A, Lavelle P (1988) Système de culture et faune des sols: quelques données. In: Feller C (Ed.) *Fertilité des sols dans les agricultures paysannes caribéennes: effet des restitutions organiques*. ORSTOM, Martinique, 85–96.
- Bartz MLC, Costa AC, Tormena CA, Souza Júnior IG, Brown GG (2010) Sobrevivência, produção e atributos químicos de coprólitos de duas espécies de minhocas (*Pontoscolex corethrurus*: Glossoscolecidae e *Amyntas gracilis*: Megascolecidae) em solos sob diferentes sistemas de manejo. *Acta Zoológica Mexicana (nueva série)* 26: 261–280.
- Bartz MLC, Pasini A, Brown GG (2013) Earthworms as soil quality indicators in Brazilian no-tillage systems. *Applied Soil Ecology* 69(1): 39–48. <https://doi.org/10.1016/j.apsoil.2013.01.011>
- Blakemore RJ (2002) *Cosmopolitan Earthworms – An Eco-Taxonomic Guide to the Peregrine Species of the World*. VermEcology, Kippax, 426 pp.
- Block W, Banage WB (1968) Population density and biomass of earthworms in some Uganda soils. *Revue d'Écologie et Biologie du Sol* 3: 515–521.
- Braga LPP, Yoshiura CA, Borges CD, Horn MA, Brown GG, Drake HL, Tsai SM (2015) Disentangling the influence of earthworms in sugarcane rhizosphere. *Scientific Reports* 6: e38923. <https://doi.org/10.1038/srep38923>

- Brown GG, James SW (2007) Ecologia, biodiversidade e biogeografia das minhocas no Brasil. In: Brown GG, Fragoso C (Eds) *Minhocas na América Latina: Biodiversidade e ecologia*. Embrapa Soja, Londrina, 297–381.
- Brown GG, Domínguez J (2010) Uso das minhocas como bioindicadoras ambientais: princípios e práticas – o 3º encontro latino americano de ecologia e taxonomia de oligoquetas (ELAETAO3). *Acta Zoológica Mexicana* (nueva série) 26: 1–18. <https://doi.org/10.21829/azm.2010.262874>
- Brown GG, Pashanasi B, Villenave C, Patron JC, Senapati BK, Giri S, Barois I, Lavelle P, Blanchart E, Blakemore RJ, Spain AV, Boyer J (1999) Effects of earthworms on plant production in the tropics. In: Lavelle P, Brussaard L, Hendrix P (Eds) *Earthworm management in tropical agroecosystems*. CABI, Wallingford, 87–147.
- Brown GG, Barois I, Lavelle P (2000) Regulation of soil organic matter dynamics and microbial activity in the drilosphere and the role of interactions with other edaphic functional domains. *European Journal of Soil Biology* 36(1): 177–198. [https://doi.org/10.1016/S1164-5563\(00\)01062-1](https://doi.org/10.1016/S1164-5563(00)01062-1)
- Brown GG, Maschio W, Froufe LCM (2009) Macrofauna do solo em sistemas agroflorestais e Mata Atlântica em regeneração nos Municípios de Barra do Turvo, SP, e Adrianópolis, PR. Embrapa Florestas, Documentos No. 184, Colombo, 51 pp.
- Brown GG, James SW, Pasini A, Nunes DH, Benito NP, Martins PT, Sautter KD (2006) Exotic, peregrine, and invasive earthworms in Brazil: diversity, distribution and effects on soils and plants. *Caribbean Journal of Science* 42: 111–117.
- Brown GG, Callaham MA, Niva CC, Feijoo A, Sautter KD, James SW, Fragoso C, Pasini A, Schmelz RM (2013) Terrestrial oligochaete research in Latin America: The importance of the Latin American Meetings on Oligochaete Ecology and Taxonomy. *Applied Soil Ecology* 69: 2–12. <https://doi.org/10.1016/j.apsoil.2012.12.006>
- Budijastuti W (2019) Type of earthworm in the banana tree habitat. *Journal of Physics: Conference Series* 1277(1): 012029. <https://doi.org/10.1088/1742-6596/1277/1/012029>
- Bueno E (2003) *Brasil: uma História* (2nd ed.). Ática, São Paulo, 480 pp.
- Bünemann EK, Bongiorno G, Bai Z, Creamer RE, De Deyn G, De Goede R, Pulleman M (2018) Soil quality—A critical review. *Soil Biology and Biochemistry* 120: 105–125.
- Burac M, Gros-Desormeaux JR, Lalubie G, Lesales T, Angin B, Breuil M, Picard R (2018) Evaluation et Suivi de la Biodiversité dans les Bananeraies: Guadeloupe et Martinique. Cihense, France, 183 pp. <https://doi.org/10.1016/j.soilbio.2018.01.030>
- Campbell CW (2018) Tropical fruits and nuts. In: Martin FW (Ed.) *Handbook of Tropical Food Crops*. CRC Press, Boca Raton, 235–274.
- Capowiez Y, Samartino S, Cadoux S, Bouchant P, Richard G, Boizard H (2012) Role of earthworms in regenerating soil structure after compaction in reduced tillage systems. *Soil Biology and Biochemistry* 55: 93–103. <https://doi.org/10.1016/j.soilbio.2012.06.013>
- Castillo FX, Vera LO (2000) Comparación de la biodiversidad de la macrofauna de suelos bananeros con manejo convencional y orgánico en EARTH. Dissertation, Guácimo, Costa Rica: EARTH University.
- CEPAGRI (2018) Centro de Pesquisas Meteorológicas e Climáticas Aplicadas a Agricultura, Clima dos municípios paulistas. <https://www.cpa.unicamp.br/outras-informacoes/clima-dos-municipios-paulistas.html>

- CILAGRO (2018) Centro Integrado de Informações Agrometeorológicas, Monitoramento Climático. <http://www.ciiagro.sp.gov.br/ciiagroonline/Listagens/MonClim/MonClimAtualEDR.asp>
- Clermont-Dauphin C, Cabidoche YM, Meynard JM (2004) Effects of intensive monocropping of bananas on properties of volcanic soils in the uplands of the French West Indies. *Soil Use and Management* 2(2): 105–113. <https://doi.org/10.1079/SUM2003231>
- Coelho GC (2017) Ecosystem services in Brazilian's southern agroforestry systems. *Tropical and Subtropical Agroecosystems* 20(3): 475–492.
- Cordeiro GPL, Amorim M, Ronquim CC (2017) Mudança de uso e ocupação da terra no município de registro, SP, entre os anos de 1987 e 2017. Embrapa Territorial. 11º Congresso Interinstitucional de Iniciação Científica, Campinas, July 2017, Instituto Agrônômico (IAC), Campinas, 10 pp.
- Cordeiro ZJM, Matos AD, Meissner Filho PE (2004) Doenças e métodos de controle. In: Borges AL, da Silva Souza L (Eds) *O cultivo da bananeira*. Cruz das Almas: Embrapa Mandioca e Fruticultura, 146–182.
- Cornwell E (2014) Effects of different agricultural systems on soil quality in Northern Limón province, Costa Rica. *Revista de Biología Tropical* 62(3): 887–897. <https://doi.org/10.15517/rbt.v62i3.14062>
- Correia MEF, Lima DA, Franco AA, Campello EFC, Tavares SRL (2001) Comunidades da macrofauna do solo em áreas de floresta secundária de mata atlântica no Estado do Rio de Janeiro. in *Anais do V Congresso de Ecologia do Brasil Ambiente x Sociedade*, 2001. Sociedade de Ecologia do Brasil, Porto Alegre.
- Cremonesi MV, Santos A, Rozane D, Bartz MLC, Brown GG (2020) Earthworm species in *Musa* spp. (plantain and banana) plantations worldwide. Mendeley Data, V1. <http://dx.doi.org/10.17632/p8ywsnj8c5.1>
- Csuzdi C, Razafindrakoto M, Hong Y (2017) Three new species of *Kynotus* from the central highlands of Madagascar (Clitellata, Megadrili). *European Journal of Taxonomy* 336: 1–14. <https://doi.org/10.5852/ejt.2017.336>
- da Silva RF, de Aquino AM, Mercante FM, de Fátima Guimarães M (2006) Macrofauna invertebrada do solo sob diferentes sistemas de produção em Latossolo da Região do Cerrado. *Pesquisa Agropecuária Brasileira* 41(4): 697–704. <https://doi.org/10.1590/S0100-204X2006000400022>
- Dale J, James A, Paul JY, Khanna H, Smith M, Peraza-Echeverria S, Harding R (2017) Transgenic Cavendish bananas with resistance to *Fusarium* wilt tropical race 4. *Nature Communications* 8(1): 1–8. <https://doi.org/10.1038/s41467-017-01670-6>
- Demetrio WC, Ribeiro RH, Nadolny H, Bartz MLC, Brown GG (2019) Earthworms in Brazilian no-tillage agriculture: Current status and future challenges. *European Journal of Soil Science* 71: 988–1005. <https://doi.org/10.1111/ejss.12918>
- Dhar S, Chaudhuri PS (2018) Earthworm communities in banana (*Musa paradisiaca*) and paddy (*Oryza sativa*) plantations of west Tripura, India. In: Solanki GS (Ed.) *Biodiversity Conservation: Strategies and Applications*. South Eastern Book Agencies, 303–320.
- Dlamini TC, Haynes RJ (2004) Influence of agricultural land use on the size and composition of earthworm communities in northern KwaZulu-Natal, South Africa. *Applied Soil Ecology* 27(1): 77–88. <https://doi.org/10.1016/j.apsoil.2004.02.003>

- FAO – Food and Agriculture Organization of the UN (2018) FAOSTAT Database. <http://www.fao.org/faostat/en/#data>
- Feijoo A (2007) Registros históricos y listado de las lombrices de tierra de Colombia. In: Brown GG, Fragoso C (Eds) *Minhocas na América Latina: biodiversidade e ecologia*. Embrapa Soja, Londrina, 141–153.
- Feijoo A, Zuluaga LF, Molina LJ (2018) New species and records of earthworms (Annelida, Oligochaeta) in plantain cropping systems in Colombia's coffee-growing region. *Zootaxa* 4496(1): 448–458. <https://doi.org/10.11646/zootaxa.4496.1.34>
- Fernandes JO, Uehara-Prado M, Brown GG (2010) Minhocas exóticas como indicadoras de perturbação antrópica em áreas de floresta atlântica. *Acta Zoológica Mexicana (nueva série)* 26: 211–217. <https://doi.org/10.21829/azm.2010.262889>
- Figuerola DMN (2019) Comparación de la macrofauna del suelo en agro sistemas de plátano en Santo Domingo y El Carmen. PhD thesis, Ecuador: Universidad Tecnológica Equinoccial, Santo Domingo.
- Fiuza DTF, Kusdra JF, Fiuza SDS (2012) Maize growth in soil with activity of giant earthworms *Chibui bari* (Oligochaeta: Glossoscolecidae). *Revista Brasileira de Ciência do Solo* 36(2): 359–366. <https://doi.org/10.1590/S0100-06832012000200005>
- Fragoso C, Rojas P (2007) Two new species of the earthworm genus *Balanteodrilus* (Oligochaeta: Acanthodrilidae) from Eastern Mexico. *Megadrilologica* 11(10): 107–114.
- Fusilero MA, Mangubat J, Ragas RE, Baguinon N, Taya H, Rasco-jr E (2013) Weed management systems and other factors affecting the earthworm population in a banana plantation. *European Journal of Soil Biology* 56(1): 89–94. <https://doi.org/10.1016/j.ejsobi.2013.03.002>
- Garcia BNR, Vieira TA, de Assis Oliveira F (2017) Tree and shrub diversity in agroforestry homegardens in rural community in eastern amazon. *Floresta* 47(4): 543–552. <https://doi.org/10.5380/rf.v47i4.48196>
- Gasparotto L, Pereira JCR, Hanada RE, Montarroyos AVV (2006) *Sigatoka-negra da bananeira*. Manaus: Embrapa Amazônia Ocidental, 18 pp.
- Geissen V, Peña-Peña K, Huerta E (2009) Effects of different land use on soil chemical properties, decomposition rate and earthworm communities in tropical Mexico. *Pedobiologia* 53(1): 75–86. <https://doi.org/10.1016/j.pedobi.2009.03.004>
- González G, Huang CY, Zou X, Rodríguez C (2006) Earthworm invasions in the tropics. *Biological Invasions* 8(6): 1247–1256. <https://doi.org/10.1007/s10530-006-9023-7>
- González WV, Hernández IM, Espinales SC (2015) Evaluación de la diversidad de la macrofauna en las fincas plataneras Cuerno Enano (AAB) en los municipios de León y Posoltega en el ciclo agrícola 2014. Undergraduate thesis, Nicaragua: Universidad Nacional Autónoma de Nicaragua, León.
- Guerra RT, Silva EG (1994) Estudo das comunidades de minhocas (Annelida, Oligochaeta) em alguns ambientes terrestres do Estado da Paraíba. *Revista Nordestina de Biologia* 9: 209–223
- Harvey CA, Villalobos JAG (2007) Agroforestry systems conserve species-rich but modified assemblages of tropical birds and bats. *Biodiversity and Conservation* 16(8): 2257–2292. <https://doi.org/10.1007/s10531-007-9194-2>

- Huerta E, Rodríguez-Olán J, Evia-Castillo I, Montejo-Meneses E, de la Cruz-Mondragón M, García-Hernández R (2005) La diversidad de lombrices de tierra (Annelida, Oligochaeta) en el estado de Tabasco, México. *Ecosistemas y Recursos Agropecuarios* 21(42): 73–83.
- Huerta E, Rodríguez-Olán J, Evia-Castillo I, Montejo-Meneses E, de la Cruz-Mondragón M, García-Hernández R, Uribe S (2007) Earthworms and soil properties in Tabasco, Mexico. *European Journal of Soil Biology* 43: 190–195. <https://doi.org/10.1016/j.ejsobi.2007.08.024>
- Huerta E, Gaspar-Genico JA, Jarquin-Sánchez A (2013) Biodiversity of Oligochaeta in traditional banana plantations of *Musa acuminata* in Tabasco, Mexico. In: Pavlicek T, Cardet P, Almeida MT, Pascoal C, Cássio F (Eds) *Advances in Earthworm Taxonomy VI* (Annelida: Oligochaeta). Kasperek Verlag, Germany, 107–133.
- IUSS/WRB – International Union of Soil Sciences and Word Reference Base (2015) World Reference Base for Soil Resources 2014, update 2015: International soil classification system for naming soils and creating legends for soil maps. World Soil Resources Reports No. 106. FAO, Rome, 192 pp.
- Kanmegne J (2004) Slash and burn agriculture in the humid forest zone of southern Cameroon: soil quality dynamics, improved fallow management and farmers' perceptions. PhD thesis, Wageningen University, Wageningen.
- Kumar TSH, Sreepada KS, Narayanan SP, Reynolds JW (2018) Megadrile earthworms (Annelida: Oligochaeta) around Udipi Power Corporation Limited (UPCL), Udipi District, Karnataka, South-West Coast of India. *Megadrilogica* 23(5): 79–91.
- Lafont A, Risède JM, Loranger-Merciris G, Clermont-Dauphin C, Dorel M, Rhino B, Lavelle P (2007) Effects of the earthworm *Pontoscolex corethrurus* on banana plants infected or not with the plant-parasitic nematode *Radopholus similis*. *Pedobiologia* 51(4): 311–318. <https://doi.org/10.1016/j.pedobi.2007.05.004>
- Lalthanzara H (2007) Ecological studies on earthworm population in agroforestry system of Mizoram. PhD thesis, Mizoram University, Mizoram.
- Lapied E, Lavelle P (2003) The peregrine earthworm *Pontoscolex corethrurus* in the East coast of Costa Rica. *Pedobiologia* 47(5–6): 471–474. <https://doi.org/10.1078/0031-4056-00215>
- Lavelle P (1997) Diversity of Soil fauna and ecosystem function. *Biology International* 33(1): 3–16.
- Lavelle P, Decaëns T, Aubert M, Barot S, Blouin M, Bureau F, Rossi JP (2006) Soil invertebrates and ecosystem services. *European Journal of Soil Biology* 42(1): 3–15. <https://doi.org/10.1016/j.ejsobi.2006.10.002>
- Lombardi Neto F, Moldenhauer WC (1992) Erosividade da chuva: sua distribuição e relação com as perdas de solo em Campinas (SP). *Bragantia* 51(2): 189–196. <https://doi.org/10.1590/S0006-87051992000200009>
- Loranger-Merciris G, Cabidoche YM, Deloné B, Quénéhervné P, Ozier-Lafontaine H (2012) How earthworm activities affect banana plant response to nematodes parasitism. *Applied Soil Ecology* 52: 1–8. <https://doi.org/10.1016/j.apsoil.2011.10.003>
- Malézieux E, Crozat Y, Dupraz C, Laurans M, Makowski D, Ozier-Lafontaine H, Valantin-Morison M (2009) Mixing plant species in cropping systems: concepts, tools and models: a review. In: Lichtfouse E, Navarrete M, Debaeke P, Véronique S, Alberola C (Eds)

- Sustainable agriculture. Springer, Dordrecht, 329–352. https://doi.org/10.1007/978-90-481-2666-8_22
- Mariappan V, Karthikairaj K, Isaiarasu L (2013) Relationship between earthworm abundance and soil quality of different cultivated lands in Rajapalayam, Tamilnadu. *World Applied Sciences Journal* 27(10): 1278–1281.
- Marin DH, Romero RA, Guzmán M, Sutton TB (2003) Black Sigatoka: an increasing threat to banana cultivation. *Plant Disease* 87(3): 208–222. <https://doi.org/10.1094/PDIS.2003.87.3.208>
- Martínez-Leiva MA (2002) Comunidades de oligoquetos (Annelida: Oligochaeta) en tres ecosistemas con diferente grado de perturbación en Cuba. PhD thesis, Instituto de Ecología y Sistemática, La Habana.
- Maschio W, Brown G, Seoane C, Froufe L (2010) Abundância e diversidade de minhocas em agroecossistemas da Mata Atlântica nos municípios do Litoral Paranaense-Morretes e Antonina. In *Anais do 4º Encontro Latino-Americano de Ecologia e Taxonomia de Oligoquetas (ELAETAO4): Minhocas como bioindicadoras ambientais: princípios e práticas*. Embrapa Florestas, Série Documentos No. 199, Colombo, 4 pp.
- Michaelson W (1900) *Das Tierreich Oligochaeta*. Friedländer and Sohn, Berlin, 575 pp.
- Molina JR, Feijoo AM (2017) Uso del suelo y efecto sobre propiedades químicas, macrofauna en cultivo de plátano andes centrales. *Suelos Ecuatoriales* 47(1, 2): 16–24.
- Myers N, Mittermeier RA, Mittermeier CG, Fonseca GAB, Kent J (2000) Biodiversity hotspots for conservation priorities, *Nature* 403: 853–858. <https://doi.org/10.1038/35002501>
- Nair KV, Manazhy J, Manazhy A, Reynolds J (2007) Earthworm Annelida Oligochaeta fauna of Kerala, India: 1. Some species from Thiruvananthapuram Corporation. *Megadrilologica* 11(8): 85–90.
- Norgrove L, Csuzdi C, Hauser S (2011) Effects of cropping and tree density on earthworm community composition and densities in central Cameroon. *Applied Soil Ecology* 49: 268–271. <https://doi.org/10.1016/j.apsoil.2011.05.008>
- Norgrove L, Hauser S (2014) Improving plantain (*Musa* spp. AAB) yields on smallholder farms in West and Central Africa. *Food Security* 6: 501–514. <https://doi.org/10.1007/s12571-014-0365-1>
- Nunes DH, Pasini A, Benito NP, Brown GG (2007) Minhocas como bioindicadoras da qualidade ambiental. Um estudo de caso na região de Jaguapitã, PR, Brasil. In: Brown GG, Fragoso C (Eds) *Minhocas na América Latina: biodiversidade e ecologia*. Embrapa Soja, Londrina, 467–480.
- OECD-FAO (2019) *OECD-FAO Agricultural Outlook 2019–2028*. OECD Publishing, Paris/Food and Agriculture Organization of the United Nations, Rome.
- Okwakol MJ (1994) The effect of change in land use on soil macrofauna communities in Mabira Forest, Uganda. *African Journal of Ecology* 32(4): 273–282. <https://doi.org/10.1111/j.1365-2028.1994.tb00578.x>
- Oliveira MAF, Maniesi V, Teixeira W, Daitx EC (2002) Caracterização isotópica de metabasitos e anfíbolitos dos grupos Açungui e Setuva na porção sul da faixa Ribeira. *Geologia USP Série Científica* 2(1): 161–170. <https://doi.org/10.5327/S1519-874X2002000100013>

- Pashanasi B (2007) Las lombrices de tierra en diferentes ecosistemas de la Amazonia Peruana. In: Brown GG, Fragoso C (Eds) *Minhocas na América Latina: biodiversidade e ecologia*. Embrapa Soja, Londrina, 207–214.
- Paul C, Griess VC, Havardi-Burger N, Weber M (2015) Timber-based agrisilviculture improves financial viability of hardwood plantations: a case study from Panama. *Agroforestry Systems* 89(2): 217–235. <https://doi.org/10.1007/s10457-014-9755-9>
- Peixoto RDG, Marochi AI (1996) A influência da minhoca *Pheretima* sp. nas propriedades de um latossolo vermelho escuro álico e no desenvolvimento de culturas em sistema de plantio direto, em Arapoti-PR. *Revista Plantio Direto* 35: 23–25.
- Plisko JD (2001) Notes on the occurrence of the introduced earthworm *Pontoscolex corethrurus* (Müller, 1857) in South Africa (Oligochaeta: Glossoscolecidae). *African Invertebrates* 42(1): 323–334.
- Price NS (1995) The origin and development of banana and plantain cultivation. In: Gowen SR (Ed.) *Bananas and Plantains*. Springer, London, 14 pp. https://doi.org/10.1007/978-94-011-0737-2_1
- Quintero EIQ (2010) Insumos e indicadores biológicos em agrossistemas com bananeiras. PhD thesis, Universidade Federal Rural do Rio de Janeiro, Seropédica.
- Razafindrakoto M, Csuzdi C, James S, Blanchart E (2017) New earthworms from Madagascar with key to the *Kynotus* species (Oligochaeta: Kynotidae). *Zoologischer Anzeiger* 268: 126–135. <https://doi.org/10.1016/j.jcz.2016.08.001>
- Reynolds JW, Fragoso C (2004) The earthworms (Oligochaeta: Acanthodrilidae, Eudrilidae, Glossoscolecidae, Lumbricidae, Megascolecidae and Ocnerodrilidae) of Bermuda. *Megadrilologica* 10(4): 17–26.
- Reynolds JW, Julka JM, Khan MN (1995) Additional earthworm records from Bangladesh (Oligochaeta: Glossoscolecidae, Megascolecidae, Moniligastridae, Ocnerodrilidae and Octochaetidae). *Megadrilologica* 6(6): 51–64.
- Ribeiro M, Metzger J, Martensen A, Ponzoni F, Hirota M (2009) The Brazilian Atlantic Forest: How much is left, and how is the remaining forest distributed? Implications for conservation. *Biological Conservation* 142: 1141–1153. <https://doi.org/10.1016/j.biocon.2009.02.021>
- Righi G (1984) *Pontoscolex* (Oligochaeta, Glossoscolecidae), a new evaluation. *Studies on Neotropical Fauna and Environment* 19(3): 159–177. <https://doi.org/10.1080/01650528409360653>
- Righi G (1988) Uma coleção de Oligochaeta da Amazônia Brasileira. *Papéis Avulsos de Zoologia* (São Paulo) 36(30): 337–351.
- Righi G (1990) *Minhocas de Mato Grosso e Rondônia*. Relatório de Pesquisa No. 12. Programa Polonoroeste. CNPq/AED, Brasília, 157 pp.
- Römbke J, Schmidt P, Höfer H (2009) The earthworm fauna of regenerating forests and anthropogenic habitats in the coastal region of Paraná. *Pesquisa Agropecuária Brasileira* 44(8): 1040–1049. <https://doi.org/10.1590/S0100-204X2009000800037>
- Salazar-Díaz R, Tixier P (2017) Effect of plant diversity on income generated by agroforestry systems in Talamanca, Costa Rica. *Agroforestry Systems* 93(2): 571–580. <https://doi.org/10.1007/s10457-017-0151-0>

- Sherlock E, Lee S, Mcphee S, Steer M, Maes JM, Csuzdi C (2011) The first earthworm collections from Nicaragua with description of two new species (Oligochaeta). *Zootaxa* 2732(1): 49–58. <https://doi.org/10.11646/zootaxa.2732.1.4>
- Siddaraju M, Sreepada KS, Krishna MP (2013) Recorded distribution of earthworms of the family Octochaetidae in Dakshina Kannada district, south west coast, Karnataka. *International Journal of Scientific and Research Publications* 3(6): 1–8.
- Siddaraju M, Sreepada KS, Reynolds JW (2010) Checklist of earthworms (Annelida: Oligochaeta) from Dakshina Kannada, Karnataka South West India. *Megadrilologica* 15(5): 65–76.
- Sims RW (1987) New species and records of earthworms from Jamaica with notes on the genus *Eutrigaster* Cognetti, 1904 (Octochaetidae: Oligochaeta). *Journal of Natural History* 21(2): 429–441. <https://doi.org/10.1080/00222938700771101>
- Sun J, Jiang JB, Qiu JP (2012) Four new species of the *Amyntas corticis*-group (Oligochaeta: Megascolecidae) from Hainan Island, China. *Zootaxa* 3458(1): 149–158. <https://doi.org/10.11646/zootaxa.3458.1.8>
- Taheri S, Pelosi C, Dupont L (2018) Harmful or useful? A case study of the exotic peregrine earthworm morphospecies *Pontoscolex corethrurus*. *Soil Biology and Biochemistry* 116(1): 277–289. <https://doi.org/10.1016/j.soilbio.2017.10.030>
- Talavera JA (1990a) Considerations about *Ocnerodrilus occidentalis* (Oligochaeta: Ocnerodrilidae) in the Canary Islands. *Bonner Zoologische Beiträge* 41: 81–87.
- Talavera JA (1990b) Peregrine earthworms from the Canary Archipelago (Oligochaeta, Megascolecidae). *Bolletino di Zoologia* 57: 159–164. <https://doi.org/10.1080/11250009009355692>
- Talavera JA (1992a) Earthworms of the banana groves from Tenerife (Canary Islands). *Soil Biology and Biochemistry* 24(12): 1369–1375. [https://doi.org/10.1016/0038-0717\(92\)90120-M](https://doi.org/10.1016/0038-0717(92)90120-M)
- Talavera JA (1992b) Octochaetid earthworms of the Canary Islands. *Bonner Zoologische Beiträge* 43(2): 339–348.
- Talavera JA (1996) Madeira earthworm fauna. *Italian Journal of Zoology* 63(1): 81–86. <https://doi.org/10.1080/11250009609356111>
- Talavera JA (2007) Gomera earthworm fauna (Canary Islands). *Italian Journal of Zoology* 74(2): 203–207. <https://doi.org/10.1080/11250000701249231>
- Talavera JA (2011) New earthworm records for Macaronesia with observations on the species, ecological characteristics and colonization history. *Pedobiologia* 54(5–6): 301–308. <https://doi.org/10.1016/j.pedobi.2011.06.001>
- Talavera JA, Pérez DI (2009) Occurrence of the Genus *Microscolex* (Oligochaeta, Acanthodrilidae) at Western Canary Islands. *Bonner Zoologische Beiträge* 56(1/2): 37–41.
- Teng SK, Aziz NAA, Mustafa M, Laboh R, Ismail IS, Devi S (2016) Potential role of endogeic earthworm *Pontoscolex corethrurus* in remediating banana blood disease: a preliminary observation. *European Journal of Plant Pathology* 145(2): 321–330. <https://doi.org/10.1007/s10658-015-0846-x>
- Timm ES, Pardo LH, Coello RP, Navarrete TC, Villegas ON, Ordonez ES (2016) Identification of differentially-expressed genes in response to *Mycosphaerella fijiensis* in the resistant *Musa* accession ‘Calcutta-4’ using suppression subtractive hybridization. *PLoS ONE* 11(8): 32–41. <https://doi.org/10.1371/journal.pone.0160083>

- Tondoh JE (1994) Effet des divers modes d'utilisation sur le peuplement animal des sols dans la région de Lamto (Côte d'Ivoire). Gestion des systèmes agro-sylvo-pastoraux tropicaux. Thesis DESS, France: Université Paris XII.
- Tondoh JE (2007) Effet de la mise en culture des forêts secondaires sur les peuplements de macroinvertébrés du sol dans la zone de contact forêt-savane de Côte d'Ivoire. *Sciences & Nature* 4(2): 197–204. <https://doi.org/10.4314/scinat.v4i2.42144>
- Tsai CF, Shen HP, Tsai SC (2000) Occurrence of the exotic earthworm *Pontoscolex corethrurus* (Müller) (Glossoscolecidae: Oligochaeta) in Taiwan. *Endemic Species Research* 2: 68–73.
- Yahia E (2019) Achieving sustainable cultivation of tropical fruits. Burleigh Dodds Science Publishing, London, 664 pp. <https://doi.org/10.19103/AS.2019.0054>
- Zicsi A (2007) An annotated checklist of the earthworms of Ecuador (Oligochaeta). Earthworms from South America 42. In: Brown GG, Fragoso C (Eds) *Minhocas na América Latina: biodiversidade e ecologia*. Embrapa Soja, Londrina, 175–199.